

A HPLC-UV Method for the Quantification of Regorafenib in Tumor

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Abstract: Regorafenib has been approved for the treatment of colorectal cancer, gastrointestinal stromal tumor and hepatocellular carcinoma. High-performance liquid chromatography (HPLC) was developed and validated for determination of regorafenib in xenograft tumors. After protein precipitation with acetonitrile, regorafenib were separated using gradient elution (C18 Ultrabase column). Quantification was performed at 262 nm. Calibration curves were linear over the range 48.8-50000 ng/ml. The assay was applied to the determination of the drug in the tumor of nude mice receiving regorafenib 50 mg orally, and could be useful for therapeutic drug monitoring of regorafenib in routine clinical practice.

Keywords: Regorafenib, HPLC-UV, tumor.

1. INTRODUCTION

Regorafenib (BAY 73-4506), an oral multikinase inhibitor of angiogenic, stromal and oncogenic receptor tyrosine kinases, approved by FDA in 2012 for the treatment of patients with metastatic colorectal cancer who have been previously treated with fluoropyrimidine-, oxaliplatin- and irinotecan-based chemotherapy, an anti-VEGF therapy, and, if RAS wild-type, an anti-EGFR therapy [1, 2]. The FDA approved use of regorafenib was extended to treatment of patients with locally advanced, unresectable or metastatic gastrointestinal stromal tumor who have been previously treated with imatinib mesylate and sunitinib malate in 2013 [3] and to treatment of patients with hepatocellular carcinoma who have been previously treated with sorafenib in 2017 [4]. Regorafenib has exhibited cytostatic or cytotoxic activity in various cell lines and xenografts [5-8] and has demonstrated its activity in early phase clinical trials in adults with cancer [9, 10]. We recently reported that regorafenib could antagonize ABCB1 or ABCG2-mediated multidrug resistance, and enhance the antitumor activity of lapatinib in preclinical models of human colorectal cancer [11-13]. In clinic, the most common regorafenib-associated adverse events include hand-foot skin reaction, rash or desquamation, hypertension, stomatitis, liver abnormalities, diarrhea, fatigue and so on [14-17]. Therefore, appropriate monitoring, and management are essential to decrease the incidence, severity and duration of regorafenib-associated adverse events.

In this study, we report a validated method that allows for the sensitive determination of regorafenib in xenograft tumor. The method exhibited excellent performance in terms of high selectivity, wide linear range (48.8–50000 ng/ml) and simpleness and cost-efficient. The method was successfully applied to study the pharmacokinetics of regorafenib in tumor.

2. MATERIAL AND METHODS

2.1. Reagents and Materials

Regorafenib (A8236) from APExBIO Technology were dissolved in acetonitrile and stored at -20 °C. HPLC grade acetonitrile (75-05-8), trifluoroacetic acid (76-05-8) and C18 Ultrabase column (Athena C18, 5 µm, 120Å) (8.462571.0001) used were from ANPEL, Shanghai. The operate system was performed using Waters e2695 series HPLC with Waters 2998 Photodiode Array Detector. All data were acquired employing Empower 3 Quantitative Analysis version analyst data processing software. Balb/c nude mice were obtained from the Guangdong Medical Laboratory Animal Center and maintained with sterilized food and water.

2.2. Sample Preparation

The stock solutions of regorafenib (300 µg/ml) prepared in acetonitrile. Standard solutions of regorafenib at concentrations of 48.8 ng/ml, 195 ng/ml, 781 ng/ml, 3125 ng/ml, 12500 ng/ml and 50000 ng/ml prepared by serial dilution of regorafenib stock solution with acetonitrile. All standard solutions were kept at -20 °C. A pharmacokinetic mouse study was conducted with the purpose of showing the applicability of the assay because human samples were not yet available. Fourteen female nude mice with 5 weeks old and 14-17

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g weight were used. Every mouse was injected subcutaneously of the human colonrectal cancer HCT116 cells (3×10^6 in 100 μL of DMEM) under the right shoulder. When the subcutaneous tumors were approximately $0.5 \times 0.5 \text{ cm}^2$ (two perpendicular diameters) in size, the mice were randomized into two groups and treated with the following regimens: vehicle alone (0.5% hydroxypropyl-methylcellulose, 0.1% Tween 80/PBS; 100 μL) and regorafenib (50 mg/kg, orally) daily for 21 days. In the 21st day, four hours after the last treatment, the xenograft tumor were taking out and a small piece of tumor tissue were chopped and lysated by RIPA buffer. The lysis was quantitated to 10 mg/ml and proteins included were precipitated with two times acetonitrile and the supernatant after high-speed centrifugation was collected into injection tubes. The supernatant could be concentrated under nitrogen if the content is too low. The test for recovery was carried out by adding 100 $\mu\text{g/ml}$ standard drugs to 50 μL blank tumor lysis and then treated in the same way as samples.

2.3. HPLC Conditions

The mobile phases were as follows: mobile phase solvent A was methanol (0.1% TFA in acetonitrile v/v) and mobile phase solvent B was distilled water (0.1% TFA in water v/v). The initial mobile phase composition of 10% solvent A and 90% solvent B was maintained for 5 min. Between 5 and 10 min, the percentage of solvent A was changed to 95%. Between 15 and 20 min, the percentage of solvent A was then returned to

10%. The overall run time was 20 min. The flow rate was 1 ml/min and the injection volume was 20-50 μL .

3. RESULTS

3.1. Chromatography and Linearity

Representative chromatograms of standard drug are shown in Figure 1. Regorafenib was eluted with a gradient with solvent A (0.1% TFA in acetonitrile) and solvent B (0.1% TFA in water) at the wavelength of 262 nm and the peak occurs at 12.2 min (Figure 1A). The linearity of the method was evaluated over a concentration range of 48–50,000 ng/ml. After log process, the stand curve of regorafenib is $y = 0.927x + 5.216$ ($R^2=0.999$) (Figure 1B and 1C). The assay showed good correlation coefficients ($r^2 > 0.99$).

3.2. Recovery

Recovery tests were performed to evaluate the accuracy of the method. The extraction recoveries for regorafenib at the reduplicative concentration levels were 97.0–98.3%. The benefit chromatogram was showed in Figure 2. Higher recovery values with $97.66 \pm 0.91\%$ were achieved, which indicates that the method is efficient and rapid.

3.3. Analysis of Samples

The control blank tumor lysis (Figure 3A) has no other peaks at the same time of 12.2 min. No significant interference was observed in the

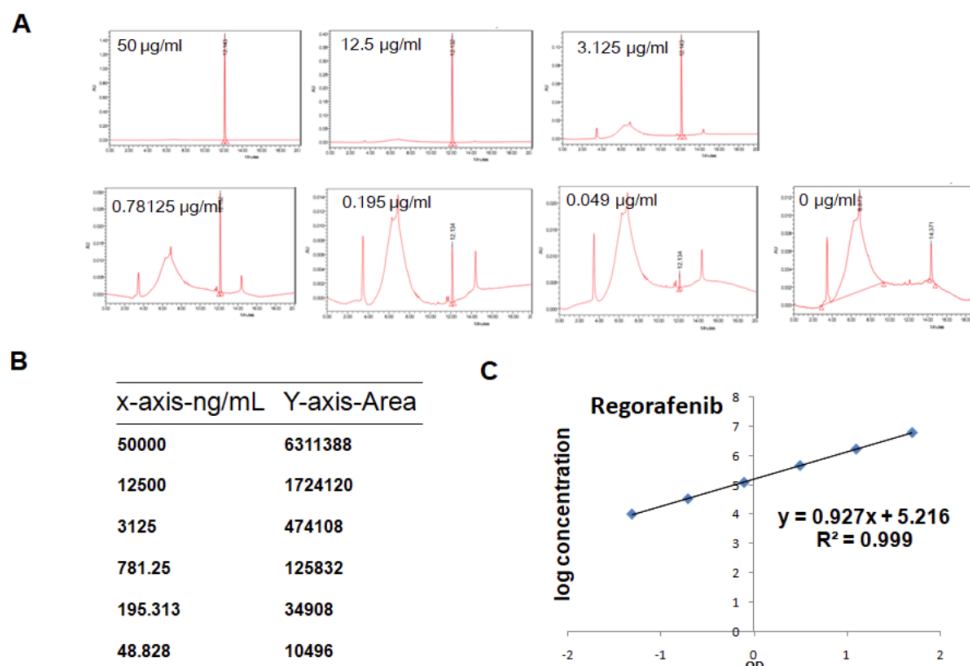


Figure 1: The representative chromatograms (A), area of peaks (B) and standard curve (C) of regorafenib are shown.

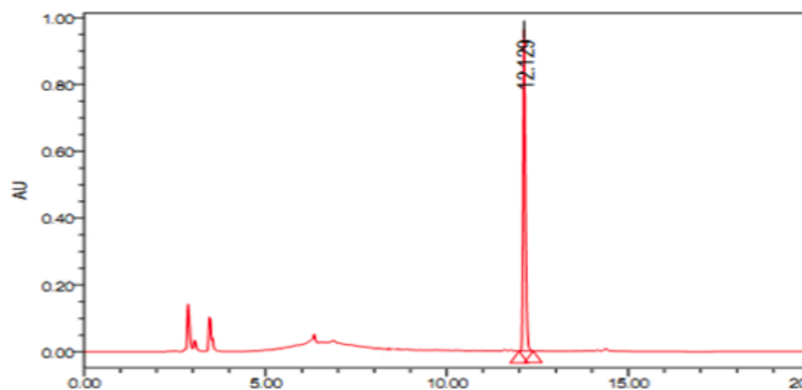


Figure 2: The representative chromatogram of regorafenib recovery test is shown.

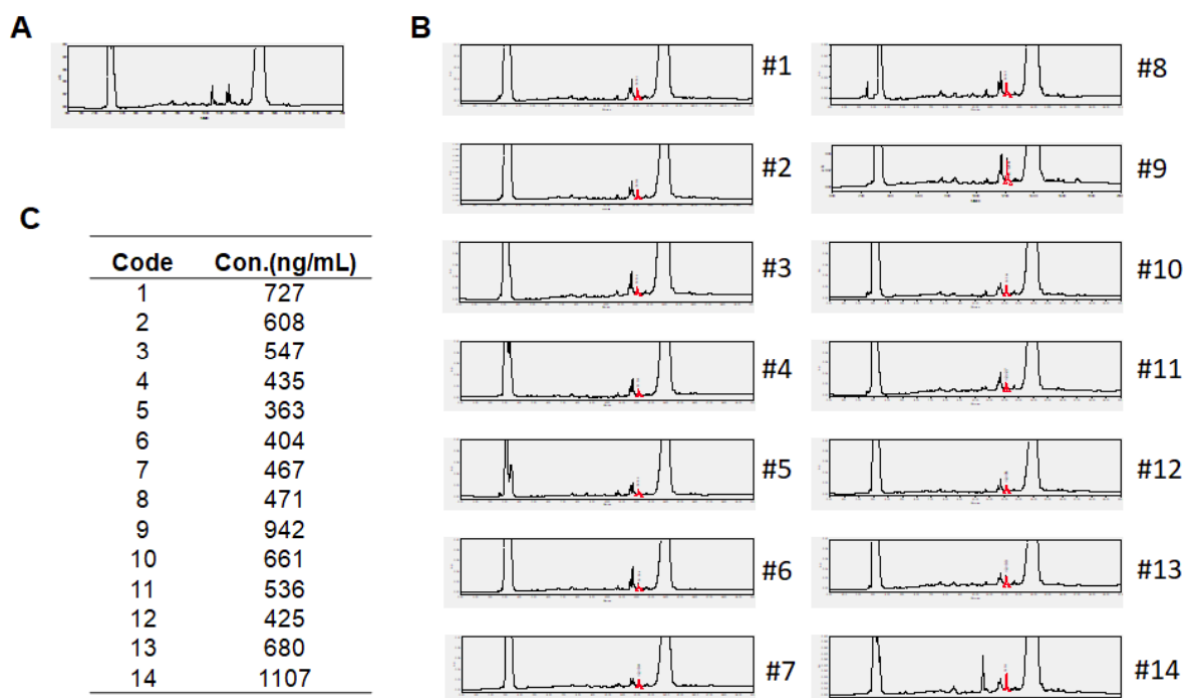


Figure 3: The chromatograms in control group (A), 14 tumor samples (B) and concentrations (C) of regorafenib are shown.

chromatograms of control tumor lysis without regorafenib treatment. Tumor regorafenib concentrations were analyzed in 14 nude mice under regorafenib monotherapy. The concentration profile of regorafenib is 598 ± 213 ng/ml, indicating that all concentrations were in the range of 48–50,000 ng/ml. The concentrations of regorafenib in tumor are demonstrated in Figure 3B and the peaks were shown in Figure 3C. Taken together, these results show that the present analytical method can be used for quantification of regorafenib in tumors.

4. DISCUSSION

Quantitative determination of regorafenib in human plasma has been reported with high-performance liquid

chromatography and ultraviolet detection, and coefficients and accuracies of variation for intra- and inter-day assays were <12.2% and <9.4% respectively for regorafenib over a linear range from 10 to 10,000 ng/mL [18]. There are two additional reported methods for analysis of regorafenib, stable-isotope dilution liquid chromatography–tandem mass spectrometry [19] and bioanalytical liquid chromatography–tandem mass spectrometric assay [20] in human plasma. Here, we established an easy and reproducible method for determination of regorafenib concentration in tumor. This assay facilitates a precious determination of regorafenib at least in the broad range of 48–50,000 ng/ml. No interference with other compounds was detected. Termovap sample concentrator is a good choice if the

drug content is beyond the lower limit of detection. It is noteworthy that the sample had better not be blown dry and then high-speed centrifugation is necessary. In the present work, runtime for 20 min linear gradient including equilibration step between two runs is a little long of this technique in the context of regorafenib in routine practice.

In conclusion, a sensitive and rapid HPLC-UV method was developed for the quantification of regorafenib in tumor, and the concentration of regorafenib in tumor were detected. This simple and cost-effective method can also be used for pharmacokinetic and pharmacodynamics studies of regorafenib and may contribute to the spreading of regorafenib monitoring in hospital laboratories not having LC/MS/MS.

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CONFLICTS OF INTEREST

There is no conflict of interest.

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