

A Marine Natural Products as Modulators of Multidrug Resistance

Tatjana P. Stanojkovic^{1,*} and Sanja Milovic²

¹*Institute of Oncology and Radiology of Serbia, Pasterova 14, 11000 Belgrade, Serbia*

²*Faculty of Pharmacy, University of Belgrade, Vojvode Stepe 450, 11221 Belgrade, Serbia*

Abstract: Multidrug resistance (MDR) which enable the tumor cells to possess intrinsic or acquired cross resistance to multiple chemotherapeutic agents simultaneously is considered to be a major challenge in cancer chemotherapy during the 21st century. numerous efflux pumps and transport proteins have been found to play important roles in MDR either the phenomenon of lowering the total intracellular retention of chemotherapeutic drugs or the redistribution of intracellular accumulation of drugs away from target organelles are two of the basic mechanisms involved in this process of MDR by transmembrane proteins which are expressed in varying concentrations in different neoplasms. Multiple compounds that have the potential to inhibit these pumps or proteins can be a future prospective for adjuvant treatment of neoplastic conditions. In this regard, compounds derived from natural products bear the advantages of low-cost and relative nontoxicity thus providing a great pool of lead structures for chemical derivatizations. This review gives an overview on chemical substances isolated from natural products of marine origin which possess the MDR modulating properties

Keywords: Multidrug resistance, Chemotherapy, ABC transporters, P-glycoprotein, MDR modulators, Cytoprotective activity, Quantitative structure-activity relationship (QASR).

INTRODUCTION

Chemotherapy is the most effective treatment for patients with cancer. However, the success of chemotherapy is seriously limited by the phenomenon of multidrug resistance (MDR) [1,2]. Anticancer drugs fail to kill cancer cells for various reasons including variations in the absorption, metabolism and delivery of the drug molecule to target tissues and tumor location in parts of the body into which the drugs do not easily penetrate [3,4,5]. Despite more than three decades of research on the subject, multidrug resistance remains one of the major obstacles to successful cancer chemotherapy [6]. This phenomenon occurs when cancer cells spontaneously become insensitive to drugs that are structurally unrelated [7]. A leading cause of MDR in cancer is the overexpression of ATP-Binding Cassette (ABC) transporters that utilize energy derived from ATP hydrolysis to actively transport anticancer drugs across biological membranes, preventing drugs from reaching their targets within a cancer cell [8,9]. Substantial efforts have been carried out to develop potent modulators of ABC drug transporters for the past two decades [10-12]. Unfortunately, these efforts have not provided successful results. The difficulty in finding an ideal inhibitor is often associated with specificity, potency and intrinsic toxicity. Adverse interactions of modulators with drugs administered in parallel or nonspecific side effects are also extremely problematic

[13]. According to previous research data, modulators targeting P-glycoprotein (P-gp)-induced MDR belong to a number of chemical classes and have been classified as the first, second and third generation of MDR reversal agents on the basis of their affinity for the transporter proteins and relative toxicity towards normal cells as marker of their side effects [11, 14, 15]. First generation modulators included drugs that were coincidentally found to be effective in sensitizing the drug resistant tumors towards chemotherapy. These include verapamil, quinine, cyclosporine A, tamoxifen and erythromycin [16,17]. The second generation modulators constituted drugs that were designed by modification of the first generation modulators and such modifications were aimed at reducing their adverse effects by eliminating their non-MDR pharmacological activities. In this group of drugs are valspodar and R-verapamil [15,18,19]. The third generation inhibitors are designed specifically for high transport affinity and low pharmacokinetic interaction [20]. These include tariquidar, biricodar, annamycin, mitotane, zosuquidar, and laniquidar. These compounds exhibit effective and potent MDR modulating activity, high affinity and selectivity for target MDR transporter(s) at low nanomolar range [21, 22]. Nevertheless, most of the agents from the first, second or third generation of MDR modulators suffer clinically from their intrinsic toxicity or from undesired effects on the pharmacokinetics of the accompanying anticancer drugs [13,23].

MDR Modulators from Natural Products

Inhibitors or modulators originating from natural sources are sometimes referred to as "Fourth

*Address correspondence to this author at the Institute of Oncology and Radiology of Serbia, Pasterova 14, 11000 Belgrade, Serbia;
E-mail: stanojkovic@ncrc.ac.rs

Generation Inhibitors". In fact, bioactive compounds from natural products provide one of the most diverse and novel chemical scaffolds suitable for the development of new inhibitors [4,24].

There is a great chemical diversity that can be utilized, as bioactive components are now extracted from plants, fungi and marine organisms, then purified and characterized [25-34]. Most importantly, natural extracts are usually low in toxicity and are well tolerated in the human body [35,36]. Accordingly, there have been significant efforts, but the progress in discovering natural MDR inhibitors is still in the early stages of exploring various extracts/active components. The main natural products that have been recognized as modulators of MDR in cancer include: flavonoids, curcumin, lignans; a variety of marine compounds including agosterol A and derivatives, siphonolol A, kendarimid A, bryostatin-1, lamellarins, ecteinascidin-743 [36-38 and references therein]. These substances belong to very different families of chemical

compounds. Some of them have very complex structures and their bioactive and anticancer activities effect different levels of tumor cell growth including blocking metabolic/enzymatic reactions, interrupting cell cycle, and direct cell killing. At least a dozen of them are in various phases of clinical trials for the treatment of cancer [39-41].

MDR Modulating Compounds Originating from Marine Organisms

Marine organisms represent a plentiful source of new bioactive compounds with promising cancer therapeutic potential, including MDR modulation properties [42-45]. A large number of distinctive secondary metabolites isolated from a wide variety of marine microorganisms, plants and invertebrates have been shown to exert anticancer effects. The most significant groups of marine compounds with anticancer properties are alkaloids, anthraquinones, benzothiazoles, macrolides, peptides, sphingolipids,

Table 1: Gives a Brief Overview of the Compounds Isolated Form the Marine Organisms that have been Undergone the Clinical Trials and Found Useful in the Treatment of Cancer in Humans

Compound name	Origin	Brand name& Manufacturer	Indication	Reference
Cytarabine	Caribbean sponge, <i>Cryptothecacrypta</i>	Cytosar-U, Ara-C (Phizer), DepoCyt (Pacira Pharmaceuticals)	Treatment of cancer and various types of leukemia	[46]
Ecteinascidin-743 (trabectedine)	Marine tunicates, <i>Ecteinascidia turbinata</i>	Yondelis® (PharmaMar)	Treatment of soft tissue sarcomas	[47]
Erbulinmesylate (synthetic form of the natural molecule halichondrin B)	Halichondrin B was isolated from marine sponge <i>Halichondria</i> sp.	Halaven® (Eisai Inc.)	Treatment of: - advanced or metastatic breast cancer; - unresectable liposarcoma	[48]
Brentuximabvedotin (Monoclonal antibody brentuximab with the monomethyl- auristatin E (MMAE), which is a synthetic analog of dolastatin-10) Dolastatin 10 was found in the sea hare <i>Dolabella auricularia</i>	Adcetris® (Seattle Genetics)	Treatment of anaplastic large T-cell systemic malignant lymphomas and Hodgkin's lymphomas	[49]
Enfortumabvedotin	Antibody, specific to nectin-4, as conjugate with MMAE	PADCEVTM® (Astellas Pharma and Seattle Genetics)	Treatment of metastatic urothelial cancer	[50]
Belantamabmafodotin	Antibody-drug conjugate with MMAE, bound to an antibody targeting B-cell maturation antigen	Blenrep® (GlaxoSmithKline)	Treatment of relapsed and refractory multiple myeloma	[51]
Plitidepsin (dehydrodidemnin B)	Marine tunicate <i>Aplidium albicans</i>	Aplidin® (PharmMar)	Treatment of leukemia, lymphoma, and multiple myeloma	[52]
Lurbinectedin	Synthetic derivative of trabectedin	Zepzelca® (PharmMar)	Treatment of metastatic small cell lung cancer	[53]

steroids, tannins, terpenes and terpenoids. Literature data point out the most significant marine sources of novel anticancer agents: sponges, coelenterates, microorganisms, algae, echinoderms, tunicates, mollusks and bryozoans [42 and references cited therein].

Among secondary metabolites of sea sponges a number of P-gp inhibitors were discovered [45]. It has been reported that a siphonane triterpene, siphonol A, isolated from the sponge *Callyspongia siphonella* efficiently reversed P-gp caused-MDR in malignant cell lines. Siphonol A increased cytotoxic effect of paclitaxel, vinblastine and colchicines in resistant malignant cell lines [52,54]. One more efficient P-gp inhibitor is polyhydroxylated sterol acetate, agosterol A, found in marine *Spongia* sp. [55-57]. Furthermore, kendarimide A isolated from the sponge *Haliclona* sp. has been shown to reverse resistance to colchicine in P-gp overexpressing KB-C2 malignant cell line [58]. Significant class of marine compounds with diverse biological and pharmacological activities including notable potential for overcoming MDR in cancer are lamellarins, polyaromatic alkaloids, which were found in *Lamellaria* sp., in ascidian, *D. chartaceum*, then in sponge, *Dendrilla* sp. and in species of unidentified ascidians [45]. Interestingly, lamellarin I exert remarkably stronger activity than verapamil in human adenocarcinoma LoVo cells resistant to doxorubicin mediated by direct inhibition of the function of P-gp pump [59]. In addition, it was reported that tetrahydroisoquinoline ecteinascidin-743, also known as trabectedin produced by chemical synthesis, originally isolated from the marine tunicate *Ecteinascidia turbinata* reversed resistance to doxorubicin and vincristine in MDR epidermal carcinoma P-gp/MDR1 overexpressing cancer cell lines [60]. Trabectedin is the marine-derived orphan drug approved for the treatment of advanced, recurrent soft tissue carcinoma in USA and Switzerland and advanced, recurrent ovarian cancer in USA and Switzerland [61]. Another effective modulator of P-gp mediated-MDR in cancer cells is bryostatin-1, a macrocyclic lactone, isolated from the marine Bryozoan *Bugula neritina*, probably a product of symbiont bacteria [62]. Moreover, marine bacteria, cyanobacteria and algae represent notable source of bioactive compounds which MDR modulation properties, such as alkaloids welwitindolinones [63] found in cyanobacteria *Hapalosiphon welwitschii*, then brominated diterpenes, parguerenes I and II derived from the Australian marine red alga *Laurencia filiformis* [64] and prenylated diketopiperazines no cardio azine A and nocardioazine B isolated from bacterium *Nocardiopsis* sp. [65].

N-Methylwelwitindolinone-cis-othiocyante, alkaloid isolated from the blue-green alga *Hapalosiphon welwitschii* were reported to reverse p-glycoprotein MDR. N-Methylwelwitindolinone C isothiocyante had MDR efficacy similar to verapamil in two tested cell lines. Also it was shown that N-methylwelwitindolinone C increased the cytotoxicity of actinomycin D and daunomycin [66]. Cyclic peptide, patellamide d, isolated from ascidian *Lissoclinum patella*, has shown cytotoxic activity and resistance in the MDR human leukemic cell line against vinblastine, adriamycin and colchicine [67]. In the marine sponge, *Discodermia dissoluta* was identified discodermolide, a polyketide, that expressed immunosuppressive and anti-tumor activities [68]. Fordiscodermolide is known that possesses the same mechanism of anti-tumor activity as taxol, and its ability to drastically decrease the MDR to taxol in ovarian carcinoma and paclitaxel-resistant colon carcinoma cell lines [69,70]. Polyoxygenated steroids, first identified in octocoral *Isis hippuris* are identified in various forms as: gorgosterol, hippuristerone, hippuristanol and hippuristerol types [71].

Many investigators tried to find the potential of well-known polysaccharide fucoidan, identified in brown algae, as the novel anticancer drug. Unfortunately, despite the cytoprotective activity of fucoidan in uveal melanoma cells and its pro-angiogenic properties, this polysaccharide showed no potential to be used as the novel medicine [72].

The extracts derived from marine bacteria family, seaweeds family and marine invertebrate superfamily, for example *Padinapavonia*, *Halimeda tuna*, *Codium bursa*, *Dysidea avara*, *Axinella cannabina*, *Achantella acuta*, *Haliclona mediterranea* extracts have shown very strong activity against human malignant cells *in vitro* [28,29,73-79]. These samples could be promising candidates for testing the ability to overcome MDR in cancer by bioactive compounds such as flavonoids, triterpenoids, quinones, lactones and sesquiterpenes.

CONCLUSION

The mentioned literature data as well as many others evidence-based data about fourth generation MDR inhibitors indicate that many of these natural products have a synergistic growth inhibitory effect with cancer drugs that are P-gp substrates including actinomycin D, puromycin, paclitaxel, vinblastine and doxorubicin [3,80,81]. Also, at the same toxicity levels the natural extracts were found to be more effective than verapamil, a standard MDR modulator, in enhancing cellular doxorubicin accumulation [82,83].

Finally, natural products represent a starting point for discovery and development of potent and effective MDR modulators, not only for their potential to be used in combination with chemotherapy treatment, but also to rationally design the semi-synthetic QSAR study analogues, with higher potency and fewer pharmacokinetic interactions.

REFERENCES

- [1] Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev Cancer* 2002; 2(1): 48-58. <https://doi.org/10.1038/nrc706>
- [2] Avendaño C, Menendez J. Inhibitors of multidrug resistance to antitumor agents (MDR). *Curr Med Chem* 2002; 9(2): 159-193. <https://doi.org/10.2174/0929867023371175>
- [3] Ullah MF. Cancer multidrug resistance (MDR): a major impediment to effective chemotherapy. *Asian Pac J Cancer Prev* 2008; 9(1): 1-6.
- [4] Wu CP, Ohnuma S, Ambudkar SV. Discovering natural product modulators to overcome multidrug resistance in cancer chemotherapy. *Curr Pharm Biotechnol* 2011; 12(4): 609-620. <https://doi.org/10.2174/138920111795163887>
- [5] Ambudkar SV, Dey S, Hrycyna CA, Ramachandra M, Pastan I, Gottesman MM. Biochemical, cellular, and pharmacological aspects of the multidrug transporter. *Annu Rev Pharmacol Toxicol* 1999; 39(1): 361-398. <https://doi.org/10.1146/annurev.pharmtox.39.1.361>
- [6] Pluchino KM, Hall MD, Goldsborough AS, Callaghan R, Gottesman MM. Collateral sensitivity as a strategy against cancer multidrug resistance. *Drug Resist Updat* 2012; 15(1-2): 98-105. <https://doi.org/10.1016/j.drug.2012.03.002>
- [7] Gottesman MM, Pastan I. Biochemistry of multidrug resistance mediated by the multidrug transporter. *Annu Rev Biochem* 1993; 62(1): 385-427. <https://doi.org/10.1146/annurev.bi.62.070193.002125>
- [8] Szakács G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev Drug Discov* 2006; 5(3): 219-234. <https://doi.org/10.1038/nrd1984>
- [9] Zhou SF. Structure, function and regulation of P-glycoprotein and its clinical relevance in drug disposition. *Xenobiotica* 2008; 38(7-8): 802-832. <https://doi.org/10.1080/00498250701867889>
- [10] Fox E, Bates SE. Tariquidar (XR9576): a P-glycoprotein drug efflux pump inhibitor. *Expert Re Anticancer Ther* 2007; 7(4): 447-459. <https://doi.org/10.1586/14737140.7.4.447>
- [11] Shukla S, Wu CP, Ambudkar SV. Development of inhibitors of ATP-binding cassette drug transporters—present status and challenges. *Expert Opin Drug Metab Toxicol* 2008; 4(2): 205-223. <https://doi.org/10.1517/17425255.4.2.205>
- [12] Pusztai L, Wagner P, Ibrahim N, Rivera E, Theriault R, Booser D, et al. Phase II study of tariquidar, a selective P-glycoprotein inhibitor, in patients with chemotherapy-resistant, advanced breast carcinoma. *Cancer* 2005; 104(4): 682-691. <https://doi.org/10.1002/cncr.21227>
- [13] Szakács G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev Drug Discov* 2006; 5(3): 219-234. <https://doi.org/10.1038/nrd1984>
- [14] Velingkar VS, Dandekar VD. Modulation of P-glycoprotein mediated multidrug resistance (MDR) in cancer using chemosensitizers. *J Pharm Sci Res* 2010; 1(2): 104-111.
- [15] Krishna R, Mayer LD. Multidrug resistance (MDR) in cancer: mechanisms, reversal using modulators of MDR and the role of MDR modulators in influencing the pharmacokinetics of anticancer drugs. *Eur J Pharm Sci* 2000; 11(4): 265-283.
- [16] Lampidis TJ, Krishan A, Planas L, Tapiero H. Reversal of intrinsic resistance to adriamycin in normal cells by verapamil. *Cancer Drug Deliv* 1986; 3(4): 251-259. <https://doi.org/10.1089/cdd.1986.3.251>
- [17] Ford JM, Hait WN. Pharmacology of drugs that alter multidrug resistance in cancer. *Pharmacol Rev* 1990; 42(3): 155-199.
- [18] Höll V, Kouba M, Dietel M, Vogt G. Stereoisomers of calcium antagonists which differ markedly in their potencies as calcium blockers are equally effective in modulating drug transport by P-glycoprotein. *Biochem Pharmacol* 1992; 43(12): 2601-2608. [https://doi.org/10.1016/0006-2952\(92\)90149-D](https://doi.org/10.1016/0006-2952(92)90149-D)
- [19] te Boekhorst PA, van Kapel J, Schoester M, Sonneveld P. Reversal of typical multidrug resistance by cyclosporin and its non-immunosuppressive analogue SDZ PSC 833 in Chinese hamster ovary cells expressing the *mdr 1* phenotype. *Cancer Chemother Pharmacol* 1992; 30(3): 238-242. <https://doi.org/10.1007/BF00686322>
- [20] Roe M, Folkes A, Ashworth P, Brumwell J, Chima L, Hunjan S, et al. Reversal of P-glycoprotein mediated multidrug resistance by novel anthranilamide derivatives. *Bioorg Med Chem Lett* 1999; 9(4): 595-600. [https://doi.org/10.1016/S0960-894X\(99\)00030-X](https://doi.org/10.1016/S0960-894X(99)00030-X)
- [21] Ullah MF. Cancer multidrug resistance (MDR): a major impediment to effective chemotherapy. *Asian Pac J Cancer Prev* 2008; 9(1): 1-6.
- [22] Liscovitch M, Lavie Y. Cancer multidrug resistance: a review of recent drug discovery research. *Drugs* 2002; 5(4): 349-355.
- [23] Limtrakul P, Siwanon S, Yodkeeree S, Duangrat C. Effect of *Stemona curtisii* root extract on P-glycoprotein and MRP-1 function in multidrug-resistant cancer cells. *Phytomedicine* 2007; 14(6): 381-389. <https://doi.org/10.1016/j.phymed.2007.03.006>
- [24] Chearwae W, Anuchapreeda S, Nandigama K, Ambudkar SV, Limtrakul P. Biochemical mechanism of modulation of human P-glycoprotein (ABCB1) by curcumin I, II, and III purified from Turmeric powder. *Biochem Pharmacol* 2004; 68(10): 2043-2052. <https://doi.org/10.1016/j.bcp.2004.07.009>
- [25] Cragg GM, Grothaus PG, Newman DJ. Impact of natural products on developing new anti-cancer agents. *Chem Rev* 2009; 109(7): 3012-3043. <https://doi.org/10.1021/cr900019j>
- [26] Newman DJ, Cragg GM. Natural products as sources of new drugs over the 30 years from 1981 to 2010. *J Nat Prod* 2012; 75(3): 311-335. <https://doi.org/10.1021/np200906s>
- [27] Hu GP, Yuan J, Sun L, She ZG, Wu JH, Lan XJ, et al. Statistical research on marine natural products based on data obtained between 1985 and 2008. *Mar Drugs* 2011; 9(4): 514-525. <https://doi.org/10.3390/md9040514>
- [28] Stanojković TP, Scaron K, Zdunić G, Kljajić Z, Grozdanić N, Antić J. In vitro antitumoral activities of *Padina pavonia* on human cervix and breast cancer cell lines. *J Med Plant Res* 2013; 7(8): 419-424.
- [29] Božić T, Novaković I, Gašić MJ, Juranić Z, Stanojković T, Tufegdžić S, et al. Synthesis and biological activity of derivatives of the marine quinone avarone. *Eur J Med Chem* 2010; 45(3): 923-929. <https://doi.org/10.1016/j.ejmech.2009.11.033>

- [30] Sipkema D, Franssen MC, Osinga R, Tramper J, Wijffels RH. Marine sponges as pharmacy. *Mar Biotechnol* 2005; 7(3): 142-162.
<https://doi.org/10.1007/s10126-004-0405-5>
- [31] Sima P, Vetvicka V. Bioactive substances with anti-neoplastic efficacy from marine invertebrates: Porifera and Coelenterata. *World J Clin Oncol* 2011; 2(11): 355-361.
<https://doi.org/10.5306/wjco.v2.i11.355>
- [32] Murti Y, Agrawal T. Marine derived pharmaceuticals-development of natural health products from marine biodiversity. *Int J Chem Tech Research* 2010; 2(4): 2198-2217.
- [33] Yu Z, Lang G, Kajahn I, Schmaljohann R, Imhoff JF, Scopularides A and B, cyclodepsipeptides from a marine sponge-derived fungus, *Scopulariopsis brevicaulis*. *J Nat Prod* 2008; 71(6): 1052-1054.
<https://doi.org/10.1021/mp070580e>
- [34] Gademann K, Sieber S. Chemical interference of biological systems with natural products. *CHIMIA* 2011; 65(11): 835-838.
<https://doi.org/10.2533/chimia.2011.835>
- [35] Patil BS, Jayaprakasha GK, Chidambara Murthy KN, Vikram A. Bioactive compounds: historical perspectives, opportunities, and challenges. *J Agric Food Chem* 2009; 57(18): 8142-8160.
<https://doi.org/10.1021/jf9000132>
- [36] Crozier A, Jaganath IB, Clifford MN. Dietary phenolics: chemistry, bioavailability and effects on health. *Nat Prod Rep* 2009; 26(8): 1001-1043.
<https://doi.org/10.1039/b802662a>
- [37] Abraham I, El Sayed K, Chen ZS, Guo H. Current status on marine products with reversal effect on cancer multidrug resistance. *Mar Drugs* 2012; 10(10): 2312-2321.
<https://doi.org/10.3390/md10102312>
- [38] Chung SY, Sung MK, Kim NH, Jang JO, Go EJ, Lee HJ. Inhibition of P-glycoprotein by natural products in human breast cancer cells. *Arch Pharm Res* 2005; 28(7): 823-828.
<https://doi.org/10.1007/BF02977349>
- [39] Haefner B. Drugs from the deep: marine natural products as drug candidates. *Drug Discov Today* 2003; 8(12): 536-544.
[https://doi.org/10.1016/S1359-6446\(03\)02713-2](https://doi.org/10.1016/S1359-6446(03)02713-2)
- [40] Garcia-Fernandez LF, Reyes F, Sanchez-Puelles JM. The marine pharmacy: new antitumoral compounds from the sea. *Pharm News* 2002; 9(6): 495-502.
- [41] Lechtenberg M, Schepmann D, Niehues M, Hellenbrand N, Wunsch B, Hensel A. Quality and functionality of saffron: quality control, species assortment and affinity of extract and isolated saffron compounds to NMDA and sigma-1 (sigma-1) receptors. *Planta Med* 2008; 74(7): 772-764.
<https://doi.org/10.1055/s-2008-1074535>
- [42] Chakraborty C, Hsu CH, Wen ZH, Lin CS. Anticancer drugs discovery and development from marine organisms. *Curr Top Med Chem* 2009; 9(16): 1536-1545.
<https://doi.org/10.2174/156802609789909803>
- [43] Sawadogo WR, Schumacher M, Teiten MH, Cerella C, Dicato M, Diederich M. A survey of marine natural compounds and their derivatives with anti-cancer activity reported in 2011. *Molecules* 2013; 18(4): 3641-3673.
<https://doi.org/10.3390/molecules18043641>
- [44] Senthilkumar K, Kim SK. Marine invertebrate natural products for anti-inflammatory and chronic diseases. *Evid Based Complement Alternat Med* 2013; 2013: 1-10.
<https://doi.org/10.1155/2013/572859>
- [45] Lopez D, Martinez-Luis S. Marine natural products with P-glycoprotein inhibitor properties. *Mar Drugs* 2014; 12(1): 525-546.
<https://doi.org/10.3390/md12010525>
- [46] Mayer AM, Glaser KB, Cuevas C, Jacobs RS, Kem W, Little RD, et al. The odyssey of marine pharmaceuticals: A current pipeline perspective. *Trends Pharmacol Sci* 2010; 31: 255-265.
<https://doi.org/10.1016/j.tips.2010.02.005>
- [47] Garcia-Carbonero R, Supko JG, Maki RG, Manola J, Ryan DP, Harmon D, et al. Ecteinascidin-743 (ET-743) for chemotherapy-naïve patients with advanced soft tissue sarcomas: multicenter phase II and pharmacokinetic study. *J Clin Oncol* 2005; 23(24): 5484-5492.
<https://doi.org/10.1200/JCO.2005.05.028>
- [48] McBride A, Butler SK. Eribulin mesylate: a novel halichondrin B analogue for the treatment of metastatic breast cancer. *Am J Health Syst Pharm* 2012; 69(9): 745-755.
<https://doi.org/10.2146/ajhp110237>
- [49] Dyshlovoy SA, Honecker F. Marine compounds and cancer: The first two decades of XXI century. *Mar Drugs* 2020; 18(1): 20-24.
<https://doi.org/10.3390/md18010020>
- [50] Hanna KS. Enfortumab vedotin to treat urothelial carcinoma. *Drugs Today (Barc)* 2020; 56(5): 329-335.
<https://doi.org/10.1358/dot.2020.56.5.3127027>
- [51] Markham A. Belantamab M. First Approval. *Drugs* 2020; 80: 1607-1613.
<https://doi.org/10.1007/s40265-020-01404-x>
- [52] Australian Public Assessment Report; The Therapeutic Goods Administration, Department of Health, Australian Government: Symonston, Australia for Plitidepsin. Available from: <https://www.tga.gov.au/sites/default/files/auspar-plitidepsin-190513.pdf>
- [53] Markham, A. Lurbinectedin: First Approval. *Drugs* 2020; 80: 1345-1353.
<https://doi.org/10.1007/s40265-020-01374-0>
- [54] Shi Z, Jain S, Kim IW, Peng XX, Abraham I, Youssef DT, et al. Siphonolol A, a marine-derived siphonolane triterpene, potentially reverses P-glycoprotein (ABCB1)-mediated multidrug resistance in cancer cells. *Cancer Sci* 2007; 98(9): 1373-1380.
<https://doi.org/10.1111/j.1349-7006.2007.00554.x>
- [55] Jain S, Abraham I, Carvalho P, Kuang YH, Shaala LA, Youssef DT, et al. Siphonolane triterpenoids: Chemistry, reversal of ABCB1/P-glycoprotein-mediated multidrug resistance, and pharmacophore modeling. *J Nat Prod* 2009; 72(7): 1291-1298.
<https://doi.org/10.1021/np900091y>
- [56] Aoki S, Yoshioka Y, Miyamoto Y, Higuchi K, Setiawan A, Murakami N, et al. Agosterol A, a novel polyhydroxylated sterol acetate reversing multidrug resistance from a marine sponge of *Spongia* sp. *Tetrahedron Lett* 1998; 39(35): 6303-6306.
[https://doi.org/10.1016/S0040-4039\(98\)01336-7](https://doi.org/10.1016/S0040-4039(98)01336-7)
- [57] Aoki S, Chen ZS, Higasiyama K, Setiawan I, Akiyama SI, Kobayashi M. Reversing effect of agosterol A, a sponge sterol acetate, on multidrug resistance in human carcinoma cells. *Jpn J Cancer Res* 2001; 92(8): 886-895.
<https://doi.org/10.1111/j.1349-7006.2001.tb01177.x>
- [58] Chen ZS, Aoki S, Komatsu M, Ueda K, Sumizawa T, Furukawa T, et al. Reversal of drug resistance mediated by multidrug resistance protein (MRP) 1 by dual effects of agosterol A on MRP1 function. *Int J Cancer* 2001; 93(1): 107-113.
<https://doi.org/10.1002/ijc.1290>
- [59] Aoki S, Cao L, Matsui K, Rachmat R, Akiyama S.I, Kobayashi M, Kendarimide A, a novel peptide reversing P-glycoprotein-mediated multidrug resistance in tumor cells, from a marine sponge of *Haliclona* sp. *Tetrahedron* 2004; 60(33): 7053-7059.
<https://doi.org/10.1016/j.tet.2003.07.020>
- [60] Quesada AR, Grávalos MG, Puentes JF. Polyaromatic alkaloids from marine invertebrates as cytotoxic compounds and inhibitors of multidrug resistance caused by P-glycoprotein. *Br J Cancer* 1996; 74(5): 677-682.
<https://doi.org/10.1038/bjc.1996.421>
- [61] Kanzaki A, Takebayashi Y, Ren XQ, Miyashita H, Mori S, Akiyama SI, et al. Overcoming multidrug drug resistance in

- P-glycoprotein/MDR1-overexpressing cell lines by ecteinascidin 743. *Mol Cancer Ther* 2002; 1(14): 1327-1334.
- [62] Carter NJ, Keam SJ. A Review of its Use in Soft Tissue Sarcoma and Ovarian Cancer. *Trabectedin. Drugs* 2010; 70(3): 355-376.
<https://doi.org/10.2165/11202860-000000000-00000>
- [63] Spitaler M, Utz I, Hilbe W, Hofmann J, Grunicke H. PKC-independent modulation of multidrug resistance in cells with mutant (V185) but not wild-type (G185) P-glycoprotein by bryostatin 1. *Biochem Pharmacol* 1998; 56(7): 861-869.
[https://doi.org/10.1016/S0006-2952\(98\)00107-5](https://doi.org/10.1016/S0006-2952(98)00107-5)
- [64] Smith CD, Zilfou JT, Stratmann K, Patterson GM, Moore RE. Welwitindolinone analogues that reverse P-glycoprotein-mediated multiple drug resistance. *Mol Pharmacol* 1995; 47(2): 241-247.
- [65] Huang XC, Sun YL, Salim AA, Chen ZS, Capon RJ. Parguerenes: Marine red alga bromoditerpenes as inhibitors of P-glycoprotein (ABCB1) in multidrug resistant human cancer cells. *Biochem Pharmacol* 2013; 85(9): 1257-1268.
<https://doi.org/10.1016/j.bcp.2013.02.005>
- [66] Raju R, Piggott AM, Huang XC, Capon RJ. Nocardioazines: A novel bridged diketopiperazine scaffold from a marine-derived bacterium inhibits P-glycoprotein. *Org Lett* 2011; 13(10): 2770-2773.
<https://doi.org/10.1021/ol200904v>
- [67] Smith CD, Zilfou JT, Stratmann K, Patterson GM, Moore RE. Welwitindolinone analogues that reverse P-glycoprotein-mediated multiple drug resistance. *Mol Pharmacol* 1995; 47(2): 241-247.
- [68] Degnan M, Hawkins CJ, Lavin MF, McCaffrey EJ, Parry DL, Van den Brenk AL, *et al.* New cyclic peptides with cytotoxic activity from the ascidian *Lissoclinum patella*. *J Med Chem* 1989; 32(6): 1349-1354.
<https://doi.org/10.1021/jm00126a034>
- [69] Gunasekera SP, Gunasekera M, Longley RE, Schulte GK. Discodermolide: a new bioactive polyhydroxylated lactone from the marine sponge *Discodermia dissoluta*. *J Org Chem* 1990; 55(16): 4912-4915.
<https://doi.org/10.1021/jo00303a029>
- [70] Kalesse M. The chemistry and biology of discodermolide. *ChemBioChem* 2000; 1(3): 171-175.
[https://doi.org/10.1002/1439-7633\(20001002\)1:3<171::AID-CBIC171>3.0.CO;2-D](https://doi.org/10.1002/1439-7633(20001002)1:3<171::AID-CBIC171>3.0.CO;2-D)
- [71] Kowalski RJ, Giannakakou P, Gunasekera SP, Longley RE, Day BW, Hamel E. The microtubule-stabilizing agent discodermolide competitively inhibits the binding of paclitaxel (Taxol) to tubulin polymers, enhances tubulin nucleation reactions more potently than paclitaxel, and inhibits the growth of paclitaxel-resistant cells. *Mol Pharmacol* 1997; 52(4): 613-622.
<https://doi.org/10.1124/mol.52.4.613>
- [72] Dithmer M, Kirsch AM, Richert E, Fuchs S, Wang F, Schmidt H, *et al.* Fucoidan does not exert anti-tumorigenic effects on uveal melanoma cell lines. *Mar Drugs* 2017; 15(7): 193-207.
<https://doi.org/10.3390/md15070193>
- [73] Chen WH, Wang SK, Duh CY. Polyhydroxylated steroids from the bamboo coral *Isis hippuris*. *Mar Drugs* 2011; 9(10): 1829-1839.
<https://doi.org/10.3390/md9101829>
- [74] Kljajić Z, Dogović N, Gašić MJ. Sterols in adriatic sea ascidians. *Comp Biochem Physiol* 1983; 75(3): 519-521.
[https://doi.org/10.1016/0305-0491\(83\)90369-3](https://doi.org/10.1016/0305-0491(83)90369-3)
- [75] Muller WEG, Diehl-Seifert B, Sobel C, Bechtold A, Kljajić Z, Dorn A. Sponge secondary metabolites: biochemical and ultrastructural localization of the antimetabolic agent avarol in *Dysidea avara*. *J Histochem Cytochem* (1986); 34(12): 1687-1690.
<https://doi.org/10.1177/34.12.3782777>
- [76] Müller WEG, Sobel C, Sachsse W, Diehl-Seifert B, Zahn RK, Eich E, *et al.* Biphasic and differential effects of the cytostatic agents avarone and avarol on DNA metabolism of human and murine T and B lymphocytes. *Eur J Cancer Clin Oncol* 1986; 22(4): 473-476.
[https://doi.org/10.1016/0277-5379\(86\)90114-8](https://doi.org/10.1016/0277-5379(86)90114-8)
- [77] Kreuter MH, Bernd A, Holzmann H, Müller-Klieser W, Maidhof A, Weissmann N, *et al.* Cytostatic activity of aeropylsinin-1 against lymphoma and epithelioma cells. *Naturforschung* 1989; 44(7-8): 680-688.
<https://doi.org/10.1515/znc-1989-7-822>
- [78] Kreuter MH, Robitzki A, Chang S, Steffen R, Michaelis M, Kljajić Z, *et al.* Production of the cytostatic agent aeropylsinin by the sponge *Verongia aerophoba* in in vitro culture. *Comp Biochem Physiol C Toxicol Pharmacol* 1992; 101(1): 183-187.
[https://doi.org/10.1016/0742-8413\(92\)90217-U](https://doi.org/10.1016/0742-8413(92)90217-U)
- [79] Pajic I, Kljajić Z, Dogović N, Sladić D, Juranic Z, Gasic MJ. A novel lectin from the sponge *Haliciona cratera*: isolation, characterization and biological activity. *Comp Biochem Physiol C Toxicol Pharmacol* 2002; 132(2): 213-221.
[https://doi.org/10.1016/S1532-0456\(02\)00068-6](https://doi.org/10.1016/S1532-0456(02)00068-6)
- [80] Fong WF, Wang C, Zhu GY, Leung CH, Yang MS, Cheung HY. Reversal of multidrug resistance in cancer cells by *Rhizoma Alismatis* extract. *Phytomedicine* 2007; 14(2-3): 160-165.
<https://doi.org/10.1016/j.phymed.2006.03.004>
- [81] Engi H, Vasas A, Redei D, Molnár J, Hohmann J. New MDR modulators and apoptosis inducers from *Euphorbia* species. *Anticancer Research* 2007; 27(5A): 3451-3458.
- [82] Bansal T, Jaggi M, Khar R, Talegaonkar S. Emerging significance of flavonoids as P-glycoprotein inhibitors in cancer chemotherapy. *J Pharm Pharmacol Sci* 2009; 12(1): 46-78.
<https://doi.org/10.18433/J3RC77>
- [83] Abraham I, El Sayed K, Chen ZS, Guo H. Current status on marine products with reversal effect on cancer multidrug resistance. *Mar Drugs* 2012; 10(10): 2312-2321.
<https://doi.org/10.3390/md10102312>

Received on 05-12-2020

Accepted on 27-12-2020

Published on 31-12-2020

DOI: <https://doi.org/10.30683/1929-2279.2020.09.11>

© 2020 Stanojkovic and Milovic; Licensee Neoplasia Research.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.