

# ABC Transporters: Maintenance of the Cancer Stem Cell Phenotype

Wei Zhang and Li-Wu Fu\*

State Key Laboratory of Oncology in Southern China, Cancer Center, Sun Yat-Sen University, Guangzhou, 510060, China

**Abstract:** The poor therapeutic response to anti-cancer treatment and inferior prognosis of carcinoma primarily result from cancer stem cells (CSCs), which initiate and maintain tumors. Recent studies have demonstrated that the molecular phenotype of CSCs mainly consists of multidrug resistance (MDR), self-renewal, multi-lineage differentiation potential (pluripotency) and tumorigenicity. Intriguingly, ATP-binding cassette (ABC) membrane transporters are highly expressed in CSCs compared to non-CSCs, and recent evidence has highlighted a link between ABC transporters and the CSC phenotype. Understanding the relationship between CSCs and ABC transporters is important as this could lead to the development of more efficacious treatment regimens. Thus, in this article, we will mainly review the relationships between ABC transporters and the phenotype of CSCs.

**Keywords:** ABC transporters, cancer stem-like cells (CSCs), multidrug resistance (MDR), self-renewal, pluripotency, side population (SP), tumorigenicity.

## INTRODUCTION

In recent years, although great progress has been made in the treatment of against cancer, the refractory status of cancer, including chemo-resistance, recurrence and metastasis, remains a major challenge. The understanding of the causes responsible for the failure of anticancer agents, recurrence and metastasis of tumors is of great importance. Emerging studies have shown that cancer stem-like cells (CSCs) play a key role in these processes [1-6]. Malignant stem-like cells have been identified in various solid tumor and leukemia. Similar to normal stem cells, CSCs are able to self-renew, differentiate, and proliferate extensively [7, 8]. The cancer mass that originates from rare stem-like cells can transfer the disease to immunodeficient mice, suggesting that these cancer stem-like cells (CSCs) are responsible for relapse following conventional or targeted cancer therapy and that eradication of CSCs may be necessary to cure the disease permanently. However, current therapeutic strategies may not effectively ablate the CSCs, leaving the potential for disease progression or relapse.

The ATP binding cassette (ABC) transporters are ubiquitous membrane proteins, consisting of both transmembrane domains (TMDs) and distinctive nucleotide-binding domains (NBDs), which generate energy from ATP hydrolysis to actively transport a variety of compounds across the membrane [9-11]. Based on sequence homology and domain

organization of the TMDs, the ABC family of transporters can be divided into seven distinct subfamilies (ABCA-ABCG), which are further divided into sub-subfamilies (except the ABCE/OABP family) [12, 13] (Table 1).

One of the important physiological roles of most of these proteins is to pump out diverse endogenous substrates including sugars, amino acids, peptides, proteins, and other hydrophobic compounds using the energy of ATP hydrolysis [10, 11]. In addition, these transporters catalyze the efflux of numerous xenobiotics, including antineoplastic drugs, thereby protecting normal tissues from cytotoxic effects [14, 15].

The overexpression of ABC drug transporters confers cross-resistance to multiple drugs belonging to different chemical classes *via* active efflux and thus, reducing the intracellular levels below that required to produce a therapeutic effect, resulting in MDR [16]. About 13~14 out of 49 members of the ABC protein family may produce MDR in cancer cells. The most important members in mediating MDR are ABCB1/P-gp, ABCG2/BCRP and ABCC1/MRP1 [11, 15, 17].

Currently, in addition to MDR, a large body of evidence indicates that there are some links between ABC transporters and the CSC phenotype [18, 19]. The elucidation of the relationship between ABC transporters and CSCs may provide a basis for improving therapeutic interventions against malignant cancer. Therefore, in this paper, we review the relationships between ABC transporters and the CSC properties including MDR, self-renewal, pluripotency and tumorigenicity.

\*Address correspondence to this author at the Cancer Center, Sun Yat-Sen University, Guangzhou 510060, Guangdong Province, China; Tel: +86-20-873-431-63; Fax: +86-20-873-431-70; E-mail: Fulw@mail.sysu.edu.cn

**Table 1: Classification of the Human ABC Transporter Family and Members that Determine MDR**

Subfamily	Alternative name	Number of proteins	Proteins that confer drug resistance	
			Name	Number
ABCA	ABC1	12	ABCA2	1
ABCB	MDR	11	ABCB1(P-gp)	3
			ABCB4(MDR3)	
			ABCB11(BSEP,SPGP)	
ABCC	MRP	13	ABCC1(MRP1)	9
			ABCC2-6(MRP2-6)	
			ABCC10(MRP7)	
			ABCC11(MRP8)	
			ABCC12(MRP9)	
ABCD	ALD	4		
ABCE	OABP	1		
ABCF	GCN20	3		
ABCG	White	5	ABCG2(BCRP)	1
Total		49		14

## 1. ABC TRANSPORTERS AND MDR OF CSCs

The failure of cancer chemotherapy can occur through acquired-resistance or intrinsic resistance to antineoplastic drugs [20, 21]. The nature of clinical drug resistance is multifactorial, involving alterations in drug targets, inactivation/detoxification of the drug, decreased drug uptake, increased drug efflux, and the dysregulation of apoptotic pathways [22]. The reduction of intracellular drug levels can be mediated by an increased expression of specific ATP binding cassette (ABC) transporters, which are responsible for multidrug resistance (MDR) [23]. The most extensively characterized ABC transporters that mediate MDR are ABCB1 (also known as MDR1 or P-glycoprotein), ABCC1 (also known as MRP1) and ABCG2 (also known as BCRP or MXR) [24].

### 1.1. ABCB1 and MDR of CSCs

ABCB1 (P-gp), a 170-kDa transmembrane glycoprotein from the superfamily of ATP binding cassette transporters, serves as an ATP-dependent efflux pump for a variety of chemicals, including many antineoplastic agents such as taxanes, anthracyclines and vinca alkaloids [25]. Furthermore, numerous studies indicate that the overexpression of ABCB1 is associated with multidrug resistance (MDR) [15, 26, 27].

Cancer stem-like cells are defined as “a small subset of cancer cells within a cancer that constitute a

reservoir of self-sustaining cells with the exclusive ability to self-renew and to cause the heterogeneous lineages of cancer cells that comprise the tumor” [28]. Currently,, there is considerable evidence indicating that ABCB1 transporters are overexpressed in various types of CSCs [29-36]. Interestingly, these cancer stem-like cells are resistant to a variety of conventional therapies [37]. Therefore, ABCB1 may be responsible for the MDR of CSCs, mainly through the efflux of antineoplastic drugs.

There are several mechanisms that may mediate the ABCB1-induced MDR of CSCs: (1) overexpression of ABCB1; (2) functional abnormality in ABCB1 [38, 39]; (3) the presence of subcellular P-gp [40]; (4) inhibition of cisplatin-induced caspase-3 activation [41]; (5) modulation of intracellular calcium homeostasis [42-46];

### 1.2. ABCC1 and MDR of CSCs

Apart from P-gp, multidrug-associated protein 1 (MRP1 or ABCC1) also plays a role in the development of drug resistance in the majority human cancers, including those of the lung, breast and prostate, as well as childhood neuroblastomas [47]. The ABCC1 transporter was the first member of the MRP family reported to be linked with MDR [48]. It catalyzes the efflux of antineoplastic drugs such as anthracyclines and mitoxantrone, as well as drugs conjugated to glutathione- (GSH), sulfate- or glucuronate [49].

Numerous studies have shown MRP1 is overexpressed in a variety of solid tumors and MRP1 mediates drug resistance in several types of cancers [47]. A study by Marques *et al.* found that Lucena cells resistant to chemotherapy (also expressing CD34<sup>+</sup>CD38<sup>-</sup>) significantly overexpressed ABCC1 genes [50]. Moreover, Vesuna *et al.* found the first direct evidence demonstrating that increased ABCC1 transporter levels in Twist-overexpressing cells lead to the development of chemo-resistance and development of the stem cell phenotype in these cells [51]. In addition, both *in vitro* and *in vivo* studies have reported that down-regulation of MRP1 in cancer cell lines or tumor xenografts produces significant chemosensitization to clinically relevant MRP1 substrate drugs, thereby verifying its role in MDR [52-54].

### 1.3. ABCG2 and MDR of CSCs

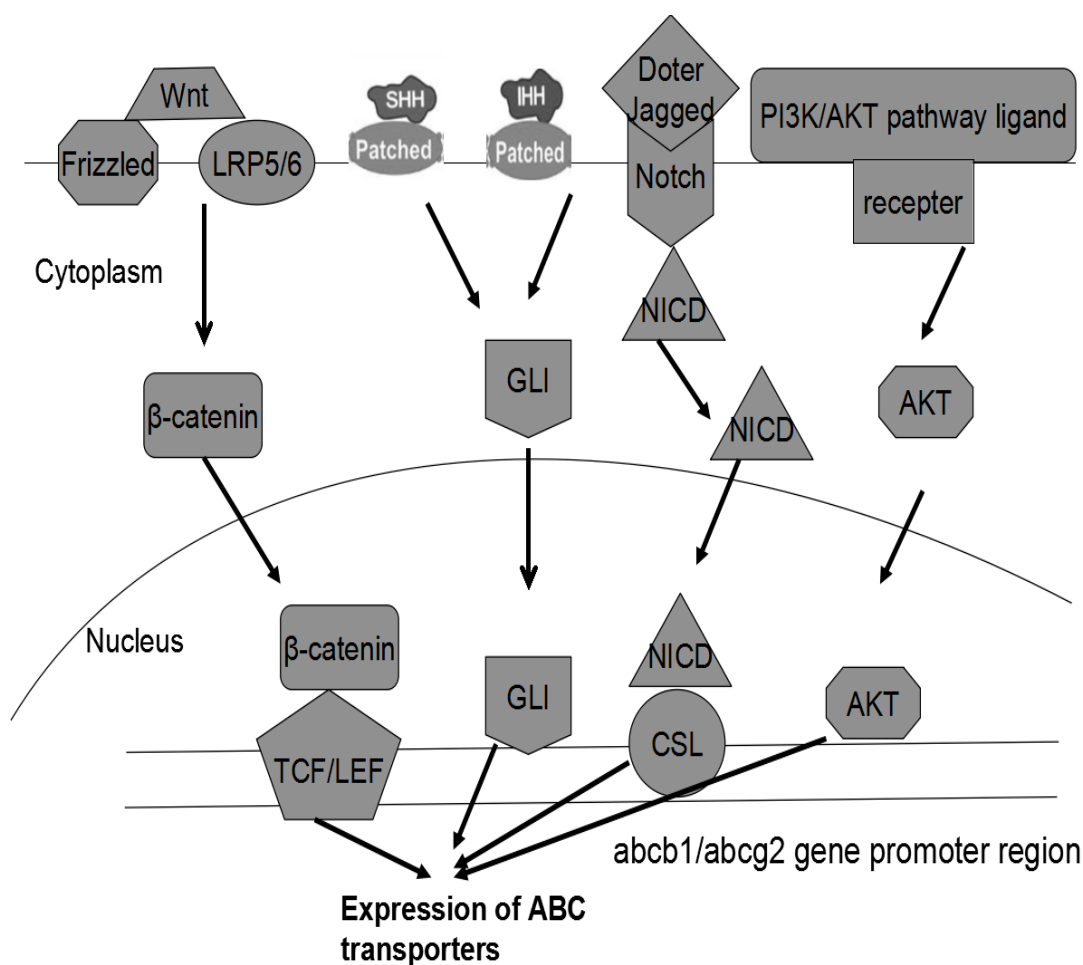
ABCG2 (BCRP, breast cancer resistance protein, ABCP or MXR) was first identified in a MCF-7/Adr-Vp3000 cancer cell line that neither expressed P-gp nor MRP1 but exhibited a high resistance to doxorubicin, mitoxantrone and daunorubicin [55]. The mediation of chemoresistance by ABCG2 was first reported by Zhou *et al.* who demonstrated that stem cells derived from ABCG2<sup>-</sup> deficient mice were more sensitive to mitoxantrone, an antineoplastic drug that is a substrate for ABCG2 [56]. Numerous *in vitro* studies indicate that the overexpression of the ABCG2 transporter is correlated with MDR in various types of cancers [57]. Furthermore, a study by Bhatia *et al.* reported that ABCG2 is expressed in the nuclear extracts of select glioblastoma and astrocytoma cell lines as well as in a human ABCG2 tumor biopsy [58].

Overall, ABCG2 and P-gp play a critical role in chemoresistance in CSCs or SP cells, though the detailed relationship between them remains unknown. ABC transporters-mediated MDR involves not only the expression level and functional activity of ABC transporters, but also the subcellular location of ABC transporters. Recent studies suggest that ABC transporter-mediated MDR may also occur by their interaction with certain cell regulatory pathways [59], down-regulation of cell surface saccharide targets [60], interaction with intracellular calcium homeostasis and by regulating endoplasmic reticulum proteins [42, 61].

## 2. ABC TRANSPORTERS AND SELF-RENEWAL CAPACITY OF CSCs

Self-renewal is the process where a stem cell divides to generate one (asymmetric division) or two

(symmetric division) daughter stem cells indistinguishable from those of the mother cells, while preserving their developmental potentials [62]. Unlike normal stem cells or embryonic stem cells, cancer stem cells have an unlimited self-renewal capability [62]. Recently, multiple lines of evidence indicate that self-renewal is regulated by a variety of signal transduction pathways, different transcription factors, and many molecular events. These include the expression of Sox-2 [63, 64], Oct-4 [65-69], Nanog [70], Myc [71], Bmi-1 [72-79] and the stem cell signaling pathways: Notch [80, 81], Wnt [82-86] and Hedgehog [87, 88]. Additionally, recent evidence indicates that transcription factors associated with self-renewal pathways modulate the expression of some ABC transporters, and thus maintain the self-renewal capability of cancer stem cells [89] (Figure 1). For example, Marques *et al.* suggested that ABCB1 may be responsible for self-renewal capacity differences between the K562 and Lucena cell line, due to the Oct-4-induced changes in P-gp expression in the Lucena cell line [50]. Also, recent *in vitro* experiments in esophageal and prostate cancer cell lines suggest that the Hh pathway increases the expression of ABC transporters P-gp and ABCG2 [90]. In addition, the knockdown of transcription factor Gli decreased the expression of BCRP and P-gp [90, 91]. Moreover, the translocation of Gli increases the expression of ABCB1, ABCC1, and ABCG2 [92]. The aforementioned studies suggest that there is an association between the aberrant self-renewal related pathways and the overexpression of ABC transporter. Currently, the specific relationship between ABC transporters and the capacity of self-renewal in CSCs remains unknown. However, there are several postulated mechanisms by which ABC transporters could produce self-renewal in CSCs. For example, it is possible that ABCG2 expression could reduce the accumulation of DNA-damaging metabolites in mouse embryonic stem cells (ESCs), which helps prevent cell differentiation but maintain self-renewal [93]. An increase in the level of ABCG2 was observed in the colony expansion of embryonic stem (ES), followed by a decrease in the level of protoporphyrin IX (PPIX) [75]. These results suggest that ABCG2 plays a role in preventing porphyrin accumulation in ES cells and excessive production of ROS from heme metabolism during colony expansion [94] (Figure 2). Overall, based on the current experimental evidence, it is possible that self-renewal-related pathways (e.g. Hedgehog) and transcription factors could modulate self-renewal process through regulating ABC transporters.



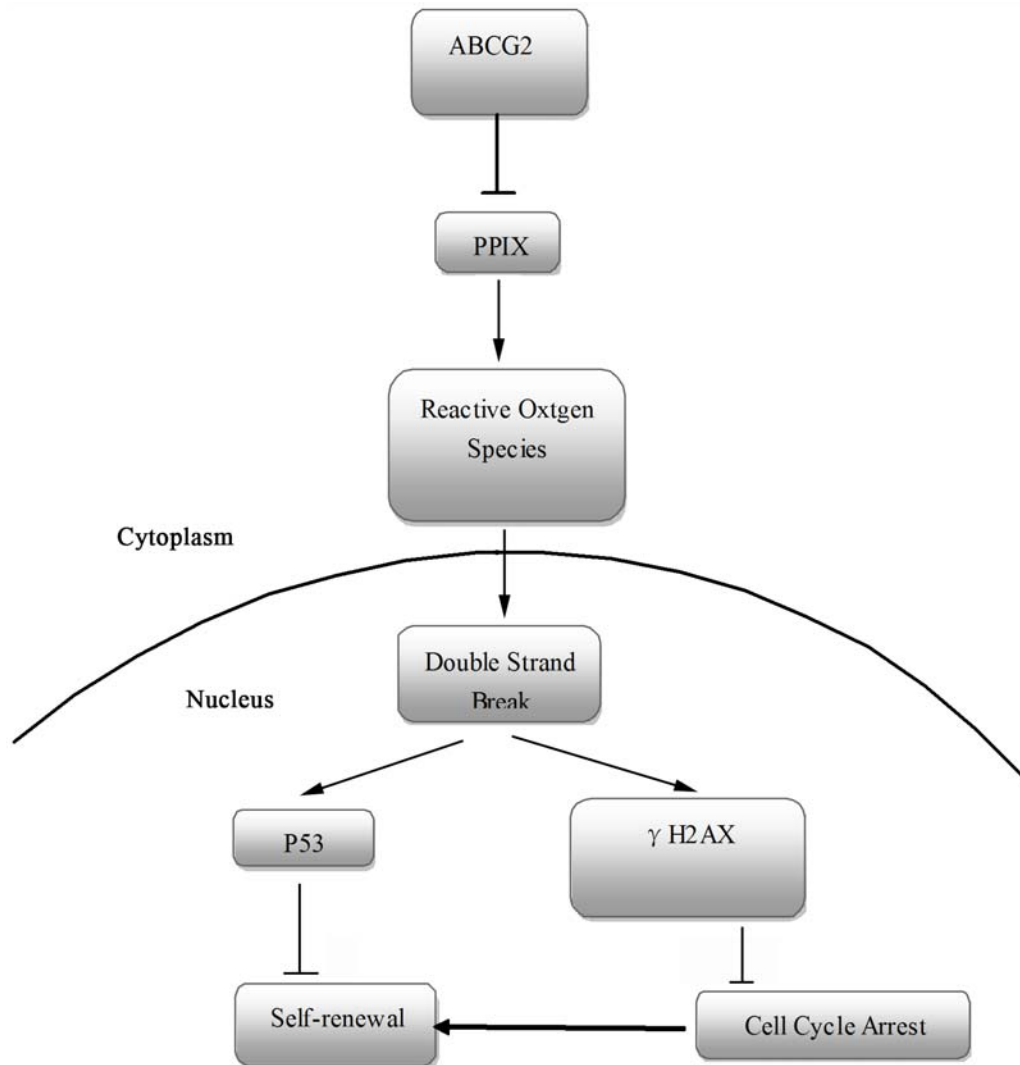
**Figure 1: The effects of CSCs associated pathways on the expression of ABC transporters.** ABC transporters have been shown to be regulated by the Hedgehog, Notch, and Wnt pathways. The activation of the Wnt pathway releases  $\beta$ -catenin from a repression complex and it translocates to the nucleus, where it interacts with the TCF/LEF consensus binding sites on the basal promoter of ABC transporter genes, which leads to their expression. Also, when the ligand SHH or IHH act with Patched, the Hedgehog pathway is activated. And its effector GLI translocates to the nucleus and bonds to the promoter of ABC transporters, which result in the expression of ABC transporters. In addition, when Doter and jagged bond to the Notch complex, an active component named NICD (Notch intracellular domain) is generated and it interacts with the CSL of promoter of the ABC transporters, thereby increasing their expression. The activation of the PI3K/AKT pathway also can produce the expression of ABC transporters through the binding of AKT with the ABC transporter gene promoter.

Therefore, the relationship between self-renewal and ABC transporters may provide a novel therapeutic strategy for overcoming the maintenance and survival of cancer stem cells.

### 3. ABC TRANSPORTERS AND PLURIPOTENCY OF CSCs

Cancer stem-like cells possess the ability to generate all types of cancer cell types (pluripotency), which form the heterogeneous macroscopic tumors [95]. Recent studies have revealed that some stem cell-like markers, such as Nanog, Oct3/4, c-Myc and Sox-2, are associated with pluripotency [68, 69, 96, 97]. Similarly, multiple lines of evidence indicate that ABC transporters, such as ABCB1 and ABCG2, are markers of cancer stem-like cells, and may be

responsible for some key traits of cancer stem-like cells, such as the self-renewal and MDR [56, 98]. However, to date, little is known about the detailed interaction of ABC transporters and pluripotency of cancer stem-like cells. Recent studies have indicated that ABC transporters may play a specific role in the maintenance of pluripotency of some types of normal tissue stem/progenitor cells, as indicated by the fluctuation in the expression of ABC transporters during differentiation and regeneration of different kinds of normal tissue stem/progenitor cells [99-101]. For example, a study by Juuti-Uusitalo K *et al.* found that the gene expression of MRP1, -3, -4, -5, and P-gp fluctuated during hESC-RPE (human stem cell-derived retinal pigment epithelial cells) maturation from undifferentiated hESC to fusiform, epithelioid, and



**Figure 2: The possible mechanism of ABCG2 mediated self-renewal of CSCs.** ABCG2 plays a role in preventing the accumulation of porphyrin and DNA damaging agents which triggers DNA damage signals such as Patm, pp53 and γH2AX.

finally to cobblestone hESC-RPE [99]. Furthermore, Islam *et al.* found that ABCB1 expression decreased during human fetal neural stem/progenitor cells (hNSPCs) differentiation and that ABCB1 may maintain hNSPCs in an undifferentiated state and could be a neural stem/progenitor marker [101]. More importantly, Barbet *et al.* reported that the expression of the 49 human ATP binding cassette (ABC) genes (except ABCB5, ABCC11, ABCC12, ABCG5, ABCG8) in pluripotent embryonic stem cells and in early- and late-stage multipotent mesenchymal stem cells may play a role in maintaining human stem cell pluripotency [100]. A recent study has shown that ABC transporters contribute to both local and global regulation of cAMP, a ubiquitous second messenger that affects multiple cell functions from maturation of the egg to cell division, growth, differentiation, and death [102]. However, the evidence discussed above is correlative and definitive

studies are needed to 1) determine whether the expression of ABC transporters is associated with the pluripotency of cancer stem cells and 2) determine the mechanism.

#### 4. ABC TRANSPORTERS AND TUMORIGENICITY OF CSCs

The ABC transporters appear to play an important role in tumorigenicity, as there are studies indicating a positive correlation between the expression of several ABC transporters and tumorigenicity (Table 2). However, there are also published studies indicating either no correlation or a negative correlation between the expression of ABC transporters and tumorigenicity (Table 2).

For example, clinical observations and genetic data from mouse models demonstrate that ABCB5 may

**Table 2: Effects of ABC Transporters on Tumorigenicity in Tumor Stem-Like Cells**

ABC Transporters	Effects on Tumorigenicity	Cancer types	Ref.
ABCB1	Positive	breast cancer, uveal melanoma	[103, 104]
ABCB5	Positive	Melanoma, colorectal cancer, oral squamous cell carcinoma	[19, 105-107]
ABCB6	Positive	hepatocellular carcinoma	[108]
ABCC4	Positive	pancreatic cancer	[109]
ABCG2	Positive	Gliomas, gallbladder cancer, ovarian cancer	[110-112]
	Negative	nasopharyngeal carcinoma	[113]
	NE	cell lines (U373 glioma and MCF7 breast cancer) and a xenograft prostate tumor (LAPC-9)	[114]

Note: ABC, ATP-binding cassette; Positive, promoting tumorigenicity; Negative, inhibiting tumorigenicity; NE, no effect on tumorigenicity.

contribute to the high tumorigenic capacity of several kinds of human cancer stem-like cells [19, 105-107]. In serial human-to-mouse xenotransplantation experiments by Tobias Schatton *et al.* [19], ABCB5<sup>+</sup> melanoma cells were shown to have a greater tumorigenic capacity than ABCB5<sup>-</sup> bulk populations. Martin Grimm *et al.* reported that ABCB5 expression in oral squamous cell carcinoma cells may be associated with tumor formation [106]. Moreover, a ABCB5<sup>+</sup> melanoma subpopulation can trigger tumorigenesis through enhanced self-renewal [107]. In addition to ABCB5, there are some studies shows that ABCC4, P-gp and ABCB6 could also promote the process of tumorigenicity of pancreatic cancer cells, breast cancer cells and uveal melanoma cells, respectively [104, 108, 109]. In general, side population cells (mainly mediated by ABCG2) always possess a higher tumorigenic capacity relative to the non-side population cells. However, an *in vivo* study by Lubna Patrawala *et al.* found that although side population cells show an increased ABCG2 mRNA expression relative to the non-side population cells, highly purified ABCG2<sup>+</sup> cancer cells have tumorigenicity similar to the ABCG2<sup>-</sup> cancer cells [114]. Therefore, the higher tumorigenicity of the side population cells may be resulted from the combined effects of several other subpopulations of cells, in addition to the ABCG2<sup>+</sup> cells. For example, cells expressing other ABC family members, such as MDR1 and MRP1 may also contribute to the cancer cell side population phenotype. However, it has been reported that the cloning efficiency of ABCG2<sup>+</sup> cells was lower than that of ABCG2<sup>-</sup> cells and unsorted cells, and the tumorigenic capacity of ABCG2<sup>+</sup> cells was also the lowest. These results may have been due to the presence of other components that enrich side populations and thus there are non-side populations in

ABCG2<sup>+</sup> cells and there are some side populations in ABCG2<sup>-</sup> cells. In addition, ABCG2<sup>+</sup> cells are not equal to side population cells and therefore do not exhibit typical properties of tumor stem-like cells.

Together, these studies indicate that the expression profile of ABC transporters may play a fundamental role in the process of tumorigenesis. However, to date, most of the evidence is largely correlative and further studies are needed to uncover the definite relationship between ABC transporters and tumorigenesis.

## 5. FUTURE DIRECTION

Currently, the clinically used antineoplastic drugs primarily target rapidly proliferating tumor cells, and have virtually no effect on cancer stem cells, thereby leading to tumor recurrence and metastasis. Therefore, the development of treatment regimens that target cancer stem cells could significantly decrease the likelihood of therapeutic failure and relapse. We have reviewed studies reporting that certain ABC transporters may play a role in maintaining the CSC phenotype. In our opinion, the effective treatment of cancer should involve the treatments that not only reduce the number of proliferating cancer cells but also ones that target CSCs. However, additional research will be required to determine the mechanism(s) by which ABC transporters mediate the self-renewal property of CSCs. We believe that as the mechanism is clarified step by step, the treatment of malignant neoplastic disease will be more effective.

## ACKNOWLEDGEMENTS

This work was supported by major science and technology project of the National Basic Research

Program (973 Program) of China (No. 2012CB967004) and National Natural Sciences Foundation of China (No. 81072669 and No. 81061160507).

## REFERENCES

- [1] O'Shaughnessy JA, *et al.* Retroviral mediated transfer of the human multidrug resistance gene (MDR-1) into hematopoietic stem cells during autologous transplantation after intensive chemotherapy for metastatic breast cancer. *Hum Gene Ther* 1994; 5(7): 891-911. <http://dx.doi.org/10.1089/hum.1994.5.7-891>
- [2] de Figueiredo-Pontes LL, *et al.* Determination of P-glycoprotein, MDR-related protein 1, breast cancer resistance protein, and lung-resistance protein expression in leukemic stem cells of acute myeloid leukemia. *Cytometry B Clin Cytom* 2008; 74(3): 163-8. <http://dx.doi.org/10.1002/cyto.b.20403>
- [3] Albarenque SM, Zwacka RM, Mohr A. Both human and mouse mesenchymal stem cells promote breast cancer metastasis. *Stem Cell Res* 2011; 7(2): 163-71. <http://dx.doi.org/10.1016/j.scr.2011.05.002>
- [4] Seol HJ, *et al.* Genetically engineered human neural stem cells with rabbit carboxyl esterase can target brain metastasis from breast cancer. *Cancer Lett* 2011; 311(2): 152-9. <http://dx.doi.org/10.1016/j.canlet.2011.07.001>
- [5] Yang L, *et al.* Gastric cancer stem-like cells possess higher capability of invasion and metastasis in association with a mesenchymal transition phenotype. *Cancer Lett* 2011; 310(1): 46-52.
- [6] Mulholland DJ, *et al.* Pten loss and RAS/MAPK activation cooperate to promote EMT and metastasis initiated from prostate cancer stem/progenitor cells. *Cancer Res* 2012; 72(7): 1878-89. <http://dx.doi.org/10.1158/0008-5472.CAN-11-3132>
- [7] Chiba T, *et al.* Cancer stem cells in hepatocellular carcinoma: Recent progress and perspective. *Cancer Lett* 2009; 286(2): 145-53. <http://dx.doi.org/10.1016/j.canlet.2009.04.027>
- [8] Al-Hajj M, Clarke MF. Self-renewal and solid tumor stem cells. *Oncogene* 2004; 23(43): 7274-82. <http://dx.doi.org/10.1038/sj.onc.1207947>
- [9] Schinkel AH, Jonker JW. Mammalian drug efflux transporters of the ATP binding cassette (ABC) family: an overview. *Adv Drug Deliv Rev* 2003; 55(1): 3-29. [http://dx.doi.org/10.1016/S0169-409X\(02\)00169-2](http://dx.doi.org/10.1016/S0169-409X(02)00169-2)
- [10] Gottesman MM, Fojo T, Bates SE. Multidrug resistance in cancer: role of ATP-dependent transporters. *Nat Rev Cancer* 2002; 2(1): 48-58. <http://dx.doi.org/10.1038/nrc706>
- [11] Glavinas H, *et al.* The role of ABC transporters in drug resistance, metabolism and toxicity. *Curr Drug Deliv* 2004; 1(1): 27-42. <http://dx.doi.org/10.2174/1567201043480036>
- [12] Vasiliou V, Vasiliou K, Nebert DW. Human ATP-binding cassette (ABC) transporter family. *Hum Genomics* 2009; 3(3): 281-90. <http://dx.doi.org/10.1186/1479-7364-3-3-281>
- [13] Dean M, Rzhetsky A, Allikmets R. The human ATP-binding cassette (ABC) transporter superfamily. *Genome Res* 2001; 11(7): 1156-66. <http://dx.doi.org/10.1101/gr.GR-1649R>
- [14] Yu M, Ocana A, Tannock IF. Reversal of ATP-binding cassette drug transporter activity to modulate chemoresistance: why has it failed to provide clinical benefit? *Cancer Metastasis Rev* 2013; 32(1-2): 211-27. <http://dx.doi.org/10.1007/s10555-012-9402-8>
- [15] Sharom FJ. ABC multidrug transporters: structure, function and role in chemoresistance. *Pharmacogenomics* 2008; 9(1): 105-27. <http://dx.doi.org/10.2217/14622416.9.1.105>
- [16] Hardwick LJ, Velamakanni S, van Veen HW. The emerging pharmacotherapeutic significance of the breast cancer resistance protein (ABCG2). *Br J Pharmacol* 2007; 151(2): 163-74. <http://dx.doi.org/10.1038/sj.bjp.0707218>
- [17] Calcagno AM, *et al.* ABC drug transporters as molecular targets for the prevention of multidrug resistance and drug-drug interactions. *Curr Drug Deliv* 2007; 4(4): 324-33. <http://dx.doi.org/10.2174/156720107782151241>
- [18] Frank NY, Frank MH. ABCB5 gene amplification in human leukemia cells. *Leuk Res* 2009; 33(10): 1303-5. <http://dx.doi.org/10.1016/j.leukres.2009.04.035>
- [19] Schatton T, *et al.* Identification of cells initiating human melanomas. *Nature* 2008; 451(7176): 345-9. <http://dx.doi.org/10.1038/nature06489>
- [20] Donnenberg VS, Donnenberg AD. Multiple drug resistance in cancer revisited: the cancer stem cell hypothesis. *J Clin Pharmacol* 2005; 45(8): 872-7. <http://dx.doi.org/10.1177/0091270005276905>
- [21] Longley DB, Johnston PG. Molecular mechanisms of drug resistance. *J Pathol* 2005; 205(2): 275-92. <http://dx.doi.org/10.1002/path.1706>
- [22] Andreeff M, Konopleva M. Mechanisms of drug resistance in AML. *Cancer Treat Res* 2002; 112: 237-62. [http://dx.doi.org/10.1007/978-1-4615-1173-1\\_12](http://dx.doi.org/10.1007/978-1-4615-1173-1_12)
- [23] Gottesman MM, Pastan I. Biochemistry of multidrug resistance mediated by the multidrug transporter. *Annu Rev Biochem* 1993; 62: 385-427. <http://dx.doi.org/10.1146/annurev.bi.62.070193.002125>
- [24] Fletcher JI, *et al.* ABC transporters in cancer: more than just drug efflux pumps. *Nat Rev Cancer* 2010; 10(2): 147-56. <http://dx.doi.org/10.1038/nrc2789>
- [25] Ambudkar SV, *et al.* Biochemical, cellular, and pharmacological aspects of the multidrug transporter. *Annu Rev Pharmacol Toxicol* 1999; 39: 361-98. <http://dx.doi.org/10.1146/annurev.pharmtox.39.1.361>
- [26] Gerlach JH, *et al.* P-glycoprotein in human sarcoma: evidence for multidrug resistance. *J Clin Oncol* 1987; 5(9): 1452-60.
- [27] Kuwazuru Y, *et al.* Expression of the multidrug transporter, P-glycoprotein, in acute leukemia cells and correlation to clinical drug resistance. *Cancer* 1990; 66(5): 868-73. [http://dx.doi.org/10.1002/1097-0142\(19900901\)66:5<868::AID-CNCR2820660510>3.0.CO;2-Z](http://dx.doi.org/10.1002/1097-0142(19900901)66:5<868::AID-CNCR2820660510>3.0.CO;2-Z)
- [28] Clarke MF, *et al.* Cancer stem cells--perspectives on current status and future directions: AACR Workshop on cancer stem cells. *Cancer Res* 2006; 66(19): 9339-44. <http://dx.doi.org/10.1158/0008-5472.CAN-06-3126>
- [29] Szotek PP, *et al.* Ovarian cancer side population defines cells with stem cell-like characteristics and Mullerian Inhibiting Substance responsiveness. *Proc Natl Acad Sci USA* 2006; 103(30): 11154-9. <http://dx.doi.org/10.1073/pnas.0603672103>
- [30] Hirschmann-Jax C, *et al.* A distinct "side population" of cells with high drug efflux capacity in human tumor cells. *Proc Natl Acad Sci USA* 2004; 101(39): 14228-33. <http://dx.doi.org/10.1073/pnas.0400067101>
- [31] Ho MM, *et al.* Side population in human lung cancer cell lines and tumors is enriched with stem-like cancer cells. *Cancer Res* 2007; 67(10): 4827-33. <http://dx.doi.org/10.1158/0008-5472.CAN-06-3557>



- [32] Wang J, *et al.* Identification of cancer stem cell-like side population cells in human nasopharyngeal carcinoma cell line. *Cancer Res* 2007; 67(8): 3716-24.  
<http://dx.doi.org/10.1158/0008-5472.CAN-06-4343>
- [33] Costello RT, *et al.* Human acute myeloid leukemia CD34+/CD38- progenitor cells have decreased sensitivity to chemotherapy and Fas-induced apoptosis, reduced immunogenicity, and impaired dendritic cell transformation capacities. *Cancer Res* 2000; 60(16): 4403-11.
- [34] Jiang X, *et al.* Chronic myeloid leukemia stem cells possess multiple unique features of resistance to BCR-ABL targeted therapies. *Leukemia* 2007; 21(5): 926-35.
- [35] Haraguchi N, *et al.* Characterization of a side population of cancer cells from human gastrointestinal system. *Stem Cells* 2006; 24(3): 506-13.  
<http://dx.doi.org/10.1634/stemcells.2005-0282>
- [36] Raaijmakers MH, *et al.* Breast cancer resistance protein in drug resistance of primitive CD34+38- cells in acute myeloid leukemia. *Clin Cancer Res* 2005; 11(6): 2436-44.  
<http://dx.doi.org/10.1158/1078-0432.CCR-04-0212>
- [37] Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. *Nat Rev Cancer* 2005; 5(4): 275-84.  
<http://dx.doi.org/10.1038/nrc1590>
- [38] Zhou WJ, *et al.* Crizotinib (PF-02341066) reverses multidrug resistance in cancer cells by inhibiting the function of P-glycoprotein. *Br J Pharmacol* 2012; 166(5): 1669-83.  
<http://dx.doi.org/10.1111/j.1476-5381.2012.01849.x>
- [39] Jovelet C, *et al.* Inhibition of P-glycoprotein functionality by vandetanib may reverse cancer cell resistance to doxorubicin. *Eur J Pharm Sci* 2012; 46(5): 484-91.  
<http://dx.doi.org/10.1016/j.ejps.2012.03.012>
- [40] Shen Y, *et al.* Mitochondrial localization of P-glycoprotein in the human breast cancer cell line MCF-7/ADM and its functional characterization. *Oncol Rep* 2012; 27(5): 1535-40.
- [41] Gibalova L, *et al.* P-glycoprotein depresses cisplatin sensitivity in L1210 cells by inhibiting cisplatin-induced caspase-3 activation. *Toxicol In Vitro* 2012; 26(3): 435-44.  
<http://dx.doi.org/10.1016/j.tiv.2012.01.014>
- [42] Sulova Z, *et al.* Does any relationship exist between P-glycoprotein-mediated multidrug resistance and intracellular calcium homeostasis. *Gen Physiol Biophys* 2009; 28 Spec No Focus: F89-95.
- [43] Barancik M, *et al.* Reversal effects of several Ca(2+)-entry blockers, neuroleptics and local anaesthetics on P-glycoprotein-mediated vincristine resistance of L1210/VCR mouse leukaemic cell line. *Drugs Exp Clin Res* 1994; 20(1): 13-8.
- [44] Witkowski JM, Miller RA. Calcium signal abnormalities in murine T lymphocytes that express the multidrug transporter P-glycoprotein. *Mech Ageing Dev* 1999; 107(2): 165-80.  
[http://dx.doi.org/10.1016/S0047-6374\(98\)00147-X](http://dx.doi.org/10.1016/S0047-6374(98)00147-X)
- [45] Gutheil JC, *et al.* Alterations in Ca2+ transport ATPase and P-glycoprotein expression can mediate resistance to thapsigargin. *J Biol Chem* 1994; 269(11): 7976-81.
- [46] Wagner-Souza K, *et al.* Resistance to thapsigargin-induced intracellular calcium mobilization in a multidrug resistant tumour cell line. *Mol Cell Biochem* 2003; 252(1-2): 109-16.  
<http://dx.doi.org/10.1023/A:1025586225941>
- [47] Munoz M, *et al.* Role of the MRP1/ABCC1 multidrug transporter protein in cancer. *IUBMB Life* 2007; 59(12): 752-7.  
<http://dx.doi.org/10.1080/15216540701736285>
- [48] Cole SP, *et al.* Overexpression of a transporter gene in a multidrug-resistant human lung cancer cell line. *Science* 1992; 258(5088): 1650-4.  
<http://dx.doi.org/10.1126/science.1360704>
- [49] Deeley RG, Cole SP. Substrate recognition and transport by multidrug resistance protein 1 (ABCC1). *FEBS Lett* 2006; 580(4): 1103-11.  
<http://dx.doi.org/10.1016/j.febslet.2005.12.036>
- [50] Marques DS, *et al.* Relationships between multidrug resistance (MDR) and stem cell markers in human chronic myeloid leukemia cell lines. *Leuk Res* 2010; 34(6): 757-62.  
<http://dx.doi.org/10.1016/j.leukres.2009.11.004>
- [51] Vesuna F, *et al.* Twist modulates breast cancer stem cells by transcriptional regulation of CD24 expression. *Neoplasia* 2009; 11(12): 1318-28.
- [52] Pajic M, *et al.* The role of the multidrug resistance-associated protein 1 gene in neuroblastoma biology and clinical outcome. *Cancer Lett* 2005; 228(1-2): 241-6.  
<http://dx.doi.org/10.1016/j.canlet.2005.01.060>
- [53] Haber M, *et al.* Altered expression of the MYCN oncogene modulates MRP gene expression and response to cytotoxic drugs in neuroblastoma cells. *Oncogene* 1999; 18(17): 2777-82.  
<http://dx.doi.org/10.1038/sj.onc.1202859>
- [54] Kuss BJ, *et al.* *In vitro* and *in vivo* downregulation of MRP1 by antisense oligonucleotides: a potential role in neuroblastoma therapy. *Int J Cancer* 2002; 98(1): 128-33.  
<http://dx.doi.org/10.1002/ijc.10159>
- [55] Doyle LA, *et al.* A multidrug resistance transporter from human MCF-7 breast cancer cells. *Proc Natl Acad Sci USA* 1998; 95(26): 15665-70.  
<http://dx.doi.org/10.1073/pnas.95.26.15665>
- [56] Zhou S, *et al.* Bcrp1 gene expression is required for normal numbers of side population stem cells in mice, and confers relative protection to mitoxantrone in hematopoietic cells *in vivo*. *Proc Natl Acad Sci USA* 2002; 99(19): 12339-44.  
<http://dx.doi.org/10.1073/pnas.192276999>
- [57] Doyle L, Ross DD. Multidrug resistance mediated by the breast cancer resistance protein BCRP (ABCG2). *Oncogene* 2003; 22(47): 7340-58.  
<http://dx.doi.org/10.1038/sj.onc.1206938>
- [58] Bhatia P, *et al.* Breast cancer resistance protein (BCRP/ABCG2) localises to the nucleus in glioblastoma multiforme cells. *Xenobiotica* 2012; 42(8): 748-55.  
<http://dx.doi.org/10.3109/00498254.2012.662726>
- [59] Lou H, Dean M. Targeted therapy for cancer stem cells: the patched pathway and ABC transporters. *Oncogene* 2007; 26(9): 1357-60.  
<http://dx.doi.org/10.1038/sj.onc.1210200>
- [60] Sulova Z, *et al.* The presence of P-glycoprotein in L1210 cells directly induces down-regulation of cell surface saccharide targets of concanavalin A. *Anticancer Res* 30(9): 3661-8.
- [61] Luquain-Costaz C, *et al.* Bis(monoacylglycero)phosphate accumulation in macrophages induces intracellular cholesterol redistribution, attenuates liver-X receptor/ATP-Binding cassette transporter A1/ATP-binding cassette transporter G1 pathway, and impairs cholesterol efflux. *Arterioscler Thromb Vasc Biol* 33(8): 1803-11.
- [62] Molofsky AV, Pardal R, Morrison SJ. Diverse mechanisms regulate stem cell self-renewal. *Curr Opin Cell Biol* 2004; 16(6): 700-7.  
<http://dx.doi.org/10.1016/j.ceb.2004.09.004>
- [63] Oesterle EC, *et al.* Sox2 and JAGGED1 expression in normal and drug-damaged adult mouse inner ear. *J Assoc Res Otolaryngol* 2008; 9(1): 65-89.  
<http://dx.doi.org/10.1007/s10162-007-0106-7>
- [64] Episkopou V. SOX2 functions in adult neural stem cells. *Trends Neurosci* 2005; 28(5): 219-21.  
<http://dx.doi.org/10.1016/j.tins.2005.03.003>
- [65] Dong Z, *et al.* Increased expression of OCT4 is associated with low differentiation and tumor recurrence in human



- hepatocellular carcinoma. *Pathol Res Pract* 2012; 208(9): 527-33.  
<http://dx.doi.org/10.1016/j.prp.2012.05.019>
- [66] Tsai CC, *et al.* Oct4 and Nanog directly regulate Dnmt1 to maintain self-renewal and undifferentiated state in mesenchymal stem cells. *Mol Cell* 2012; 47(2): 169-82.  
<http://dx.doi.org/10.1016/j.molcel.2012.06.020>
- [67] Oka M, *et al.* Differential role for transcription factor Oct4 nucleocytoplasmic dynamics in somatic cell reprogramming and self-renewal of embryonic stem cells. *J Biol Chem* 2013; 288(21): 15085-97.  
<http://dx.doi.org/10.1074/jbc.M112.448837>
- [68] Tsai CC, *et al.* Oct4 and Nanog directly regulate Dnmt1 to maintain self-renewal and undifferentiated state in mesenchymal stem cells. *Mol Cell* 47(2): 169-82.
- [69] da Cunha JM, *et al.* Pluripotent stem cell transcription factors during human odontogenesis. *Cell Tissue Res* 353(3): 435-41.
- [70] da Cunha JM, *et al.* Pluripotent stem cell transcription factors during human odontogenesis. *Cell Tissue Res* 2013; 353(3): 435-41.  
<http://dx.doi.org/10.1007/s00441-013-1658-y>
- [71] Varlakhanova NV, *et al.* myc maintains embryonic stem cell pluripotency and self-renewal. *Differentiation* 2010; 80(1): 9-19.  
<http://dx.doi.org/10.1016/j.diff.2010.05.001>
- [72] Wang Y, *et al.* Bmi-1 regulates self-renewal, proliferation and senescence of human fetal neural stem cells *in vitro*. *Neurosci Lett* 2010; 476(2): 74-8.  
<http://dx.doi.org/10.1016/j.neulet.2010.04.006>
- [73] Lukacs RU, *et al.* Bmi-1 is a crucial regulator of prostate stem cell self-renewal and malignant transformation. *Cell Stem Cell* 2010; 7(6): 682-93.  
<http://dx.doi.org/10.1016/j.stem.2010.11.013>
- [74] Raaphorst FM, Self-renewal of hematopoietic and leukemic stem cells: a central role for the Polycomb-group gene Bmi-1. *Trends Immunol* 2003; 24(10): 522-4.  
[http://dx.doi.org/10.1016/S1471-4906\(03\)00241-2](http://dx.doi.org/10.1016/S1471-4906(03)00241-2)
- [75] Iwama A, *et al.* Enhanced self-renewal of hematopoietic stem cells mediated by the polycomb gene product Bmi-1. *Immunity* 2004; 21(6): 843-51.  
<http://dx.doi.org/10.1016/j.immuni.2004.11.004>
- [76] Iwama A, *et al.* Epigenetic regulation of hematopoietic stem cell self-renewal by polycomb group genes. *Int J Hematol* 2005; 81(4): 294-300.  
<http://dx.doi.org/10.1532/IJH97.05011>
- [77] Nakauchi H, *et al.* Polycomb gene product Bmi-1 regulates stem cell self-renewal. *Ernst Schering Res Found Workshop* 2005; (54): 85-100.
- [78] Gong H, Zhang YC, Liu WL. [Regulatory effects of Bmi-1 gene on self-renewal of hematopoietic stem cells--review]. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 2006; 14(2): 413-5.
- [79] Oguro H. [Regulation of hematopoietic stem cell self-renewal by a polycomb group gene product, Bmi-1]. *Rinsho Ketsueki* 2006; 47(5): 363-70.
- [80] Balenci L, van der Kooy D. Notch signaling induces retinal stem-like properties in perinatal neural retina progenitors and promotes symmetric divisions in adult retinal stem cells. *Stem Cells Dev* 2013;
- [81] Zheng Y, *et al.* A rare population of CD24(+)ITGB4(+) Notch(hi) cells drives tumor propagation in NSCLC and requires Notch3 for self-renewal. *Cancer Cell* 2013; 24(1): 59-74.  
<http://dx.doi.org/10.1016/j.ccr.2013.05.021>
- [82] Heideel FH, Mar BG, Armstrong SA. Self-renewal related signaling in myeloid leukemia stem cells. *Int J Hematol* 2011; 94(2): 109-17.  
<http://dx.doi.org/10.1007/s12185-011-0901-0>
- [83] Cai C, Zhu X. The Wnt/beta-catenin pathway regulates self-renewal of cancer stem-like cells in human gastric cancer. *Mol Med Rep* 2012; 5(5): 1191-6.
- [84] Huang J, *et al.* Maintenance of hematopoietic stem cells through regulation of Wnt and mTOR pathways. *Nat Med* 2012; 18(12): 1778-85.  
<http://dx.doi.org/10.1038/nm.2984>
- [85] Merrill BJ. Wnt pathway regulation of embryonic stem cell self-renewal. *Cold Spring Harb Perspect Biol* 2012; 4(9): a007971;
- [86] Park JS, *et al.* Six2 and Wnt regulate self-renewal and commitment of nephron progenitors through shared gene regulatory networks. *Dev Cell* 2012; 23(3): 637-51.  
<http://dx.doi.org/10.1016/j.devcel.2012.07.008>
- [87] Nusslein-Volhard C, Wieschaus E. Mutations affecting segment number and polarity in *Drosophila*. *Nature* 1980; 287(5785): 795-801.  
<http://dx.doi.org/10.1038/287795a0>
- [88] Ingham PW, McMahon AP. Hedgehog signaling in animal development: paradigms and principles. *Genes Dev* 2001; 15(23): 3059-87.  
<http://dx.doi.org/10.1101/gad.938601>
- [89] Keshet GI, *et al.* MDR1 expression identifies human melanoma stem cells. *Biochem Biophys Res Commun* 2008; 368(4): 930-6.  
<http://dx.doi.org/10.1016/j.bbrc.2008.02.022>
- [90] Sims-Mourtada J, *et al.* Sonic Hedgehog promotes multiple drug resistance by regulation of drug transport. *Oncogene* 2007; 26(38): 5674-9.  
<http://dx.doi.org/10.1038/sj.onc.1210356>
- [91] Sims-Mourtada J, *et al.* Hedgehog: an attribute to tumor regrowth after chemoradiotherapy and a target to improve radiation response. *Clin Cancer Res* 2006; 12(21): 6565-72.  
<http://dx.doi.org/10.1158/1078-0432.CCR-06-0176>
- [92] Santisteban M. ABC transporters as molecular effectors of pancreatic oncogenic pathways: the Hedgehog-Gli model. *J Gastrointest Cancer* 2010; 41(3): 153-8.  
<http://dx.doi.org/10.1007/s12029-010-9144-1>
- [93] Zeng H, *et al.* Lack of ABCG2 expression and side population properties in human pluripotent stem cells. *Stem Cells* 2009; 27(10): 2435-45.  
<http://dx.doi.org/10.1002/stem.192>
- [94] Susanto J, *et al.* Porphyrin homeostasis maintained by ABCG2 regulates self-renewal of embryonic stem cells. *PLoS One* 2008; 3(12): e4023;
- [95] Lobo NA, *et al.* The biology of cancer stem cells. *Annu Rev Cell Dev Biol* 2007; 23: 675-99.  
<http://dx.doi.org/10.1146/annurev.cellbio.22.010305.104154>
- [96] Loh YH, *et al.* The Oct4 and Nanog transcription network regulates pluripotency in mouse embryonic stem cells. *Nat Genet* 2006; 38(4): 431-40.  
<http://dx.doi.org/10.1038/ng1760>
- [97] Masui S, *et al.* Pluripotency governed by Sox2 via regulation of Oct3/4 expression in mouse embryonic stem cells. *Nat Cell Biol* 2007; 9(6): 625-35.  
<http://dx.doi.org/10.1038/ncb1589>
- [98] Zabierowski SE, Herlyn M. Learning the ABCs of melanoma-initiating cells. *Cancer Cell* 2008; 13(3): 185-7.  
<http://dx.doi.org/10.1016/j.ccr.2008.02.015>
- [99] Juuti-Uusitalo K, *et al.* Efflux protein expression in human stem cell-derived retinal pigment epithelial cells. *PLoS One* 2012; 7(1): e30089;
- [100] Barbet R, *et al.* Expression of the 49 human ATP binding cassette (ABC) genes in pluripotent embryonic stem cells and in early- and late-stage multipotent mesenchymal stem cells: possible role of ABC plasma membrane transporters in maintaining human stem cell pluripotency. *Cell Cycle* 2012; 11(8): 1611-20.  
<http://dx.doi.org/10.4161/cc.20023>

- [101] Islam MO, *et al.* Characterization of ABC transporter ABCB1 expressed in human neural stem/progenitor cells. *FEBS Lett* 2005; 579(17): 3473-80.  
<http://dx.doi.org/10.1016/j.febslet.2005.05.019>
- [102] Cheepala S, *et al.* Cyclic nucleotide compartmentalization: contributions of phosphodiesterases and ATP-binding cassette transporters. *Annu Rev Pharmacol Toxicol* 2013; 53: 231-53.  
<http://dx.doi.org/10.1146/annurev-pharmtox-010611-134609>
- [103] Calcagno AM, *et al.* Prolonged drug selection of breast cancer cells and enrichment of cancer stem cell characteristics. *J Natl Cancer Inst* 2010; 102(21): 1637-52.  
<http://dx.doi.org/10.1093/jnci/djq361>
- [104] Landreville S, *et al.* ABCB1 identifies a subpopulation of uveal melanoma cells with high metastatic propensity. *Pigment Cell Melanoma Res* 2011; 24(3): 430-7.  
<http://dx.doi.org/10.1111/j.1755-148X.2011.00841.x>
- [105] Wilson BJ, *et al.* ABCB5 identifies a therapy-refractory tumor cell population in colorectal cancer patients. *Cancer Res* 2011; 71(15): 5307-16.  
<http://dx.doi.org/10.1158/0008-5472.CAN-11-0221>
- [106] Grimm M, *et al.* ABCB5 expression and cancer stem cell hypothesis in oral squamous cell carcinoma. *Eur J Cancer* 2012; 48(17): 3186-97.  
<http://dx.doi.org/10.1016/j.ejca.2012.05.027>
- [107] Lin JY, *et al.* Genetically determined ABCB5 functionality correlates with pigmentation phenotype and melanoma risk. *Biochem Biophys Res Commun* 2013; 436(3): 536-42.  
<http://dx.doi.org/10.1016/j.bbrc.2013.06.006>
- [108] Polireddy K, *et al.* Functional significance of the ATP-binding cassette transporter B6 in hepatocellular carcinoma. *Mol Oncol* 2011; 5(5): 410-25.  
<http://dx.doi.org/10.1016/j.molonc.2011.07.005>
- [109] Zhang Z, *et al.* The ABCC4 gene is a promising target for pancreatic cancer therapy. *Gene* 2012; 491(2): 194-9.  
<http://dx.doi.org/10.1016/j.gene.2011.09.029>
- [110] Bleau AM, *et al.* PTEN/PI3K/Akt pathway regulates the side population phenotype and ABCG2 activity in glioma tumor stem-like cells. *Cell Stem Cell* 2009; 4(3): 226-35.  
<http://dx.doi.org/10.1016/j.stem.2009.01.007>
- [111] Li XX, *et al.* Characterization of cancer stem-like cells derived from a side population of a human gallbladder carcinoma cell line, SGC-996. *Biochem Biophys Res Commun* 419(4): 728-34.
- [112] Luo LJ, *et al.* [Analysis of the characteristics of side population cells in the human ovarian cancer cell line OVCAR-3]. *Zhonghua Fu Chan Ke Za Zhi* 47(4): 281-5.
- [113] Zhang H, *et al.* Identification of ABCG2(+) cells in nasopharyngeal carcinoma cells. *Oncol Rep* 27(4): 1177-87.
- [114] Patrawala L, *et al.* Side population is enriched in tumorigenic, stem-like cancer cells, whereas ABCG2+ and ABCG2- cancer cells are similarly tumorigenic. *Cancer Res* 2005; 65(14): 6207-19.  
<http://dx.doi.org/10.1158/0008-5472.CAN-05-0592>

Received on 28-11-2013

Accepted on 13-01-2014

Published on 13-02-2014

DOI: <http://dx.doi.org/10.6000/1929-2279.2014.03.01.1>